

SITE CHARACTERIZATION SAMPLING  
FOR CONTAMINATED MATERIAL MANAGEMENT PURPOSES  
UPGRADE MAIN GATE BARRIERS AND REPAIR BOUNDARY FENCE  
TRUAX FIELD  
APRIL 10, 2020

For project XGFG 192006 soil samples will be collected at the locations discussed below. All samples will be tested for the included list of PFAS compounds. This field information will be used to develop a contaminated materials management plan. The management plan will describe the reuse or disposal of contaminated soil and/or groundwater generated during site preparation and building construction.

**SAMPLING:**

As described in the attached drawings construction of new barriers and swing gates will involve limited soil work and no soil investigation sampling will be required at these project sites. The estimated 9-20 cubic yards of soil generated during the construction the new 200 feet of security fencing in each direction from the main gate shall be stockpiled and tested for management purposes. For the planned installation of 3100 feet of new perimeter fencing soil samples shall be collected every 200 feet at depths of 1-3feet below ground surface.

No groundwater samples will be collected during construction of the security features or perimeter fencing.

**ANALYSIS**

All soil samples will be analyzed at a lab, approved by the Department, to conduct PFAS analysis. QA/QC requirements will be laboratory specific.

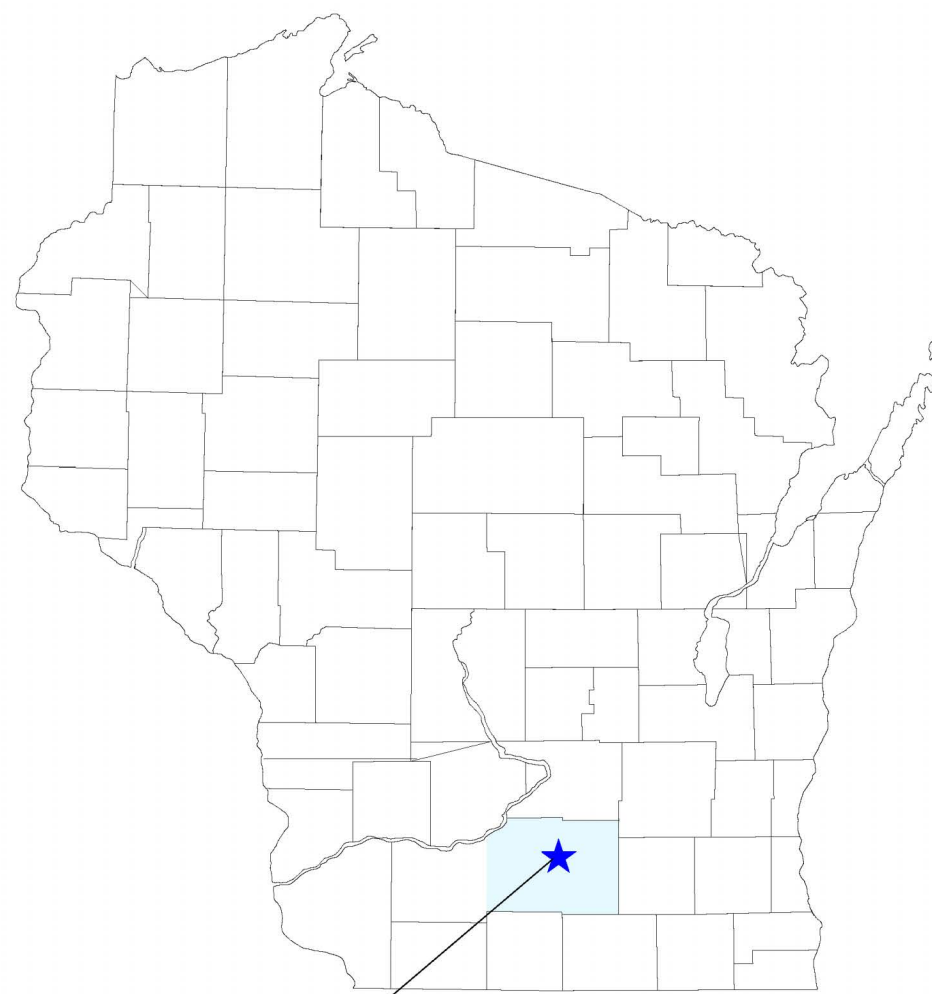
EMERGENCY CALL 911



NORTH



# PROJECT XGFG 192006



TRUAX FIELD  
MADISON, WI

WISCONSIN AIR NATIONAL GUARD  
TRUAX FIELD  
AIR NATIONAL GUARD BASE  
115<sup>TH</sup> FIGHTER WING

## UPGRADE MAIN GATE BARRIERS & REPAIR BOUNDARY FENCE

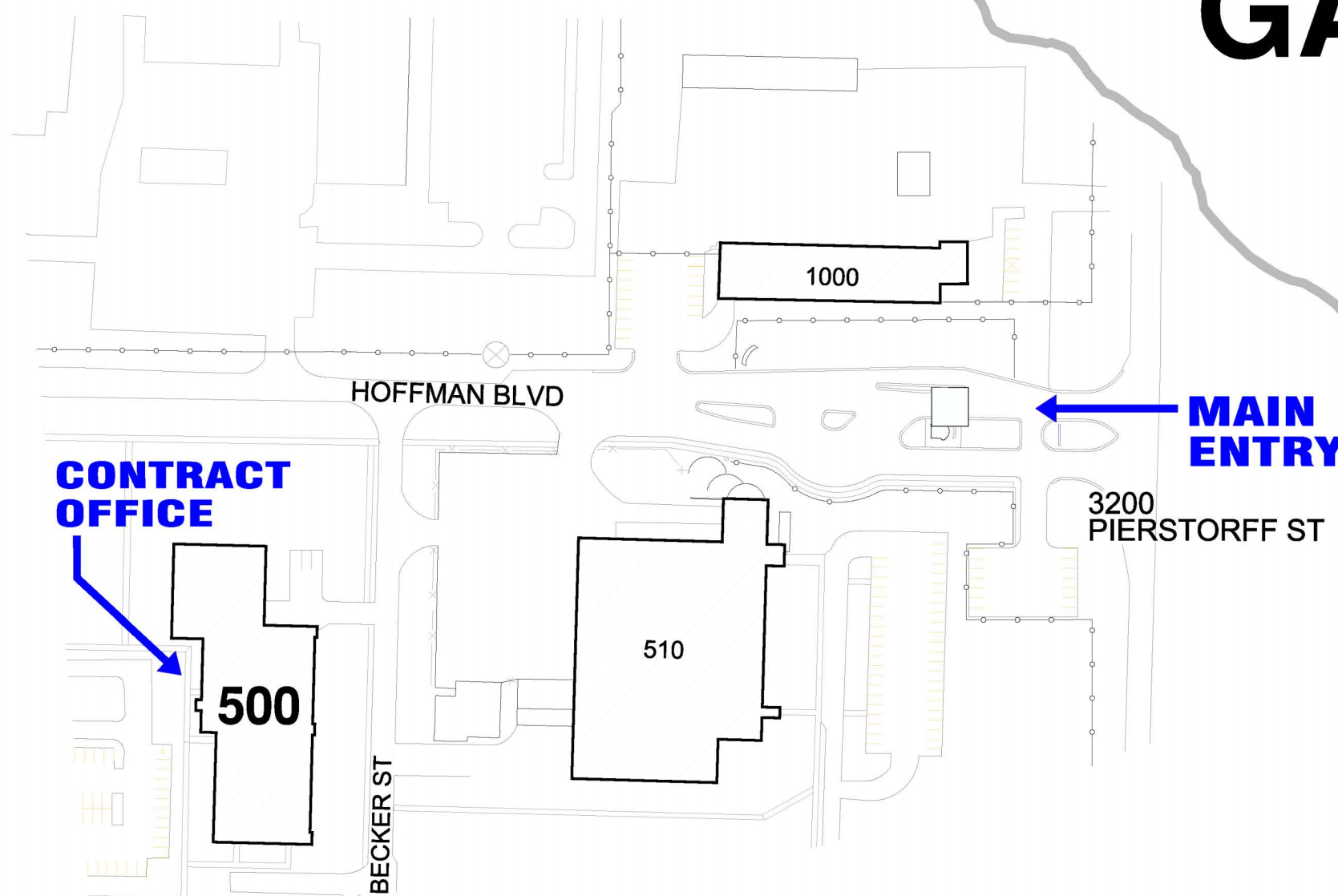
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### IMPORTANT NUMBERS

608-245-4548 CONTRACTING OFFICER  
608-245-4571 BASE ENGINEERING  
608-245-4530 SECURITY  
608-245-4561 FIRE  
608-245-4437 SAFETY

**T1**  
SHEET



### PROJECT LOCATION

115<sup>TH</sup> FIGHTER WING  
TRUAX FIELD AIR NATIONAL GUARD BASE

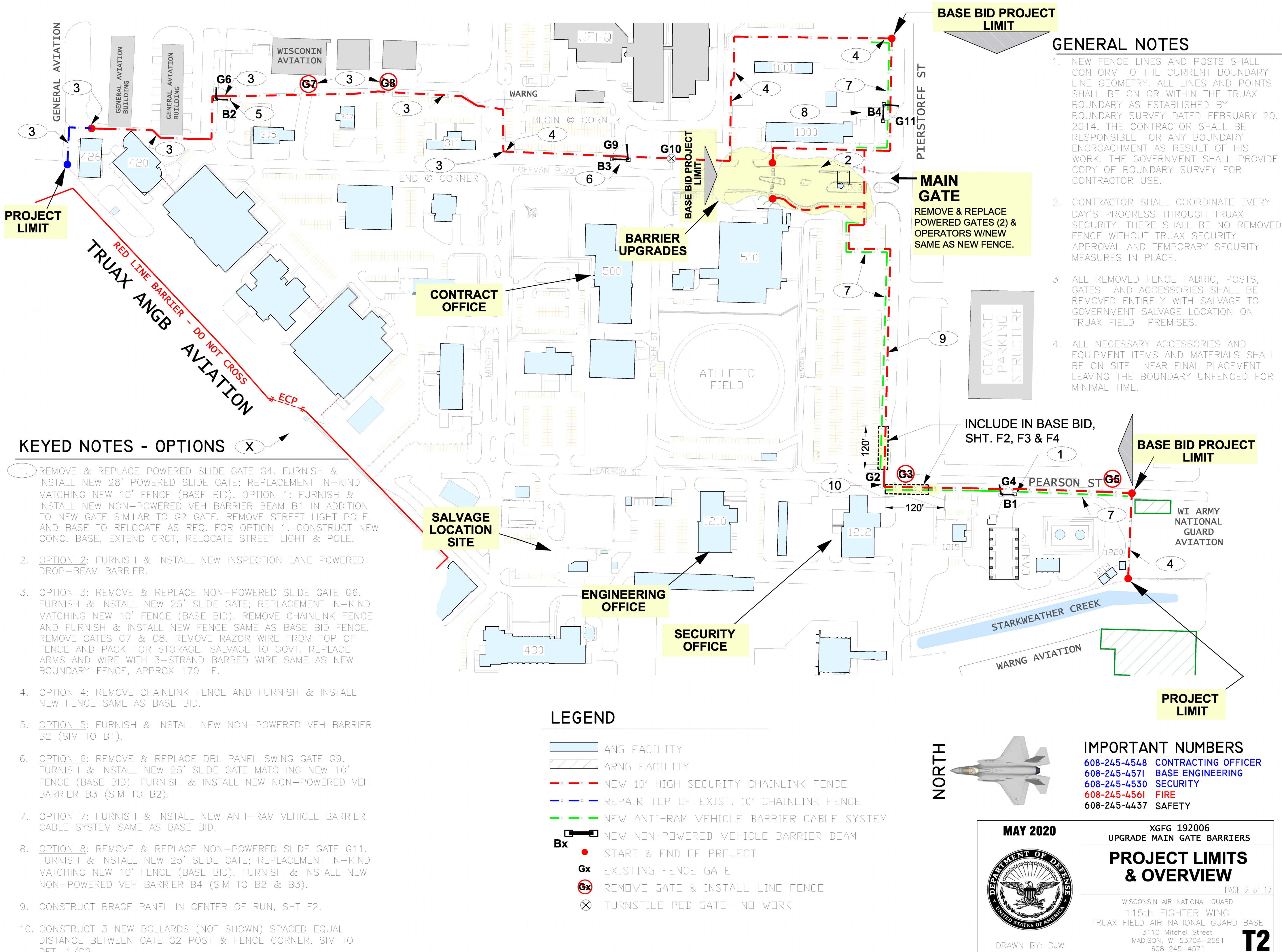
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MAY 2020

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GENERAL NOTES

1. NEW FENCE LINES AND POSTS SHALL CONFORM TO THE CURRENT BOUNDARY LINE GEOMETRY. ALL LINES AND POINTS SHALL BE ON OR WITHIN THE TRUAX BOUNDARY AS ESTABLISHED BY BOUNDARY SURVEY DATED FEBRUARY 20, 2014. THE CONTRACTOR SHALL BE RESPONSIBLE FOR ANY BOUNDARY ENCROACHMENT AS RESULT OF HIS WORK. THE GOVERNMENT SHALL PROVIDE COPY OF BOUNDARY SURVEY FOR CONTRACTOR USE.
2. CONTRACTOR SHALL COORDINATE EVERY DAY'S PROGRESS THROUGH TRUAX SECURITY. THERE SHALL BE NO REMOVED FENCE WITHOUT TRUAX SECURITY APPROVAL AND TEMPORARY SECURITY MEASURES IN PLACE.
3. ALL REMOVED FENCE FABRIC, POSTS, GATES AND ACCESSORIES SHALL BE REMOVED ENTIRELY WITH SALVAGE TO GOVERNMENT SALVAGE LOCATION ON TRUAX FIELD PREMISES.
4. ALL NECESSARY ACCESSORIES AND EQUIPMENT ITEMS AND MATERIALS SHALL BE ON SITE NEAR FINAL PLACEMENT LEAVING THE BOUNDARY UNFENCED FOR MINIMAL TIME.

KEYED NOTES - OPTIONS

1. REMOVE & REPLACE POWERED SLIDE GATE G4. FURNISH & INSTALL NEW 28' POWERED SLIDE GATE; REPLACEMENT IN-KIND MATCHING NEW 10' FENCE (BASE BID). OPTION 1: FURNISH & INSTALL NEW NON-POWERED VEH BARRIER BEAM B1 IN ADDITION TO NEW GATE SIMILAR TO G2 GATE. REMOVE STREET LIGHT POLE AND BASE TO RELOCATE AS REQ. FOR OPTION 1. CONSTRUCT NEW CONC. BASE, EXTEND CRCT, RELOCATE STREET LIGHT & POLE.
2. OPTION 2: FURNISH & INSTALL NEW INSPECTION LANE POWERED DROP-BEAM BARRIER.
3. OPTION 3: REMOVE & REPLACE NON-POWERED SLIDE GATE G6. FURNISH & INSTALL NEW 25' SLIDE GATE; REPLACEMENT IN-KIND MATCHING NEW 10' FENCE (BASE BID). REMOVE CHAINLINK FENCE AND FURNISH & INSTALL NEW FENCE SAME AS BASE BID FENCE. REMOVE GATES G7 & G8. REMOVE RAZOR WIRE FROM TOP OF FENCE AND PACK FOR STORAGE. SALVAGE TO GOVT. REPLACE ARMS AND WIRE WITH 3-STRAND BARBED WIRE SAME AS NEW BOUNDARY FENCE, APPROX 170 LF.
4. OPTION 4: REMOVE CHAINLINK FENCE AND FURNISH & INSTALL NEW FENCE SAME AS BASE BID.
5. OPTION 5: FURNISH & INSTALL NEW NON-POWERED VEH BARRIER B2 (SIM TO B1).
6. OPTION 6: REMOVE & REPLACE DBL PANEL SWING GATE G9. FURNISH & INSTALL NEW 25' SLIDE GATE MATCHING NEW 10' FENCE (BASE BID). FURNISH & INSTALL NEW NON-POWERED VEH BARRIER B3 (SIM TO B2).
7. OPTION 7: FURNISH & INSTALL NEW ANTI-RAM VEHICLE BARRIER CABLE SYSTEM SAME AS BASE BID.
8. OPTION 8: REMOVE & REPLACE NON-POWERED SLIDE GATE G11. FURNISH & INSTALL NEW 25' SLIDE GATE; REPLACEMENT IN-KIND MATCHING NEW 10' FENCE (BASE BID). FURNISH & INSTALL NEW NON-POWERED VEH BARRIER B4 (SIM TO B2 & B3).
9. CONSTRUCT BRACE PANEL IN CENTER OF RUN, SHT F2.
10. CONSTRUCT 3 NEW BOLLARDS (NOT SHOWN) SPACED EQUAL DISTANCE BETWEEN GATE G2 POST & FENCE CORNER, SIM TO DET. 1/D2.

LEGEND

- ANG FACILITY
- ARNG FACILITY
- NEW 10' HIGH SECURITY CHAINLINK FENCE
- REPAIR TOP OF EXIST. 10' CHAINLINK FENCE
- NEW ANTI-RAM VEHICLE BARRIER CABLE SYSTEM
- NEW NON-POWERED VEHICLE BARRIER BEAM
- START & END OF PROJECT
- EXISTING FENCE GATE
- REMOVE GATE & INSTALL LINE FENCE
- TURNSTILE PED GATE- NO WORK

IMPORTANT NUMBERS

- 608-245-4548 CONTRACTING OFFICER
- 608-245-4571 BASE ENGINEERING
- 608-245-4530 SECURITY
- 608-245-4561 FIRE
- 608-245-4437 SAFETY

MAY 2020



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XGFG 192006  
UPGRADE MAIN GATE BARRIERS

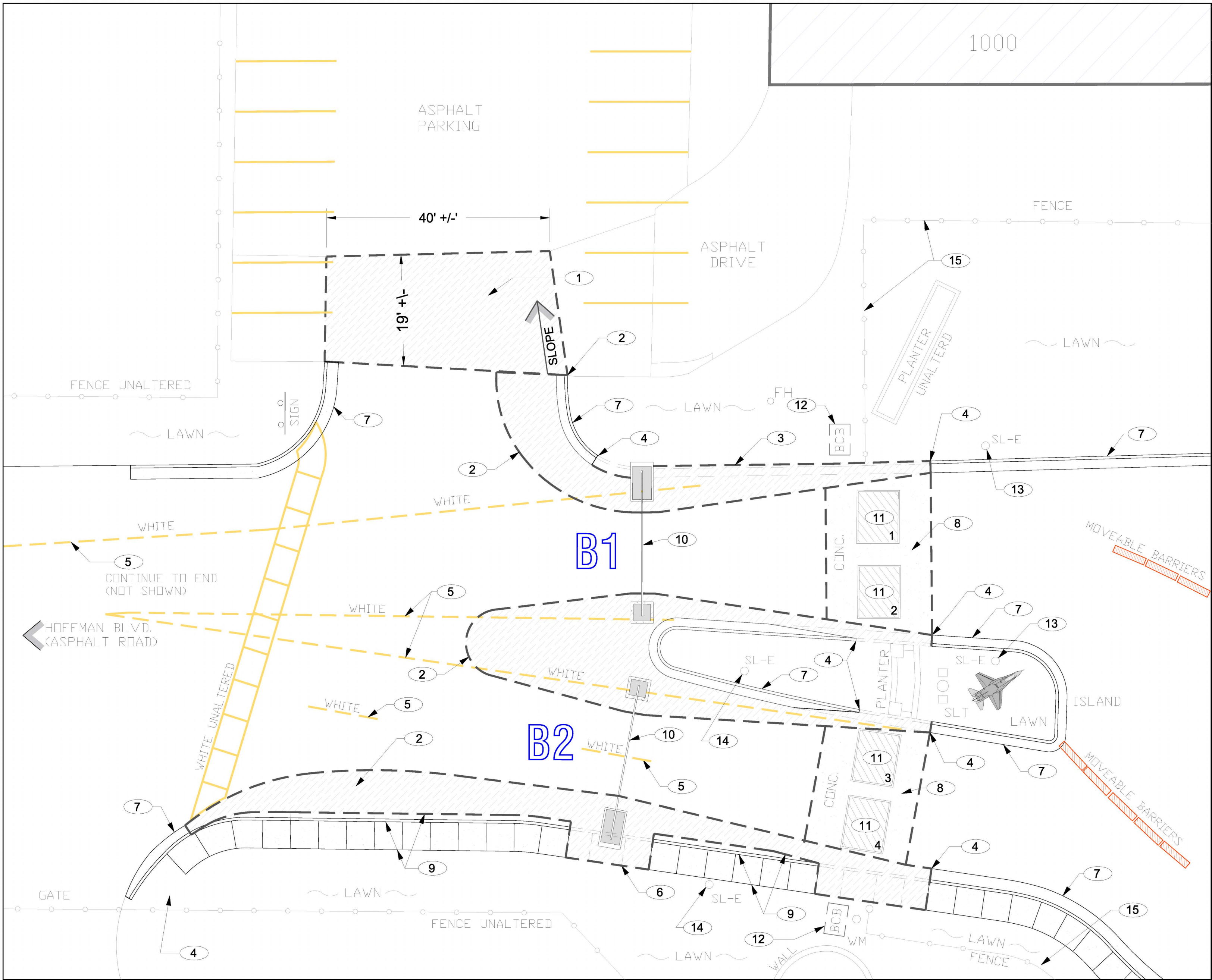
**PROJECT LIMITS  
& OVERVIEW**

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WISCONSIN AIR NATIONAL GUARD  
115th FIGHTER WING  
TRUAX FIELD AIR NATIONAL GUARD BASE  
3110 Mitchel Street  
MADISON, WI 53704-2591  
608 245-4571

**T2**





KEYED NOTES

1. NEATLY SAW-CUT & REMOVE ASPHALT (FULL DEPTH) INCLUDING BASE MATERIAL. REPLACE WITH 3.5" THICK NEW ASPHALT IN 2 EQUAL LIFTS OF WEARING COURSE AND FINISH COURSE OVER FINISH GRADED & WELL COMPACTED AGG BASE. DEPTH OF EXISTING ASPHALT IS UNKNOWN.
2. NEATLY SAW-CUT TO LINE OF NEW CONCRETE GUTTER. REMOVE ASPHALT (FULL DEPTH) INCLUDING AGGREGATE & EARTHEN SUBBASE MATERIAL AS REQ. TO CONSTRUCT NEW CURB AS DETAILED. RAISE FIN. ELEV. 6" AND TRANSITION TO EXIST. GRADE.
3. REMOVE ASPHALT (FULL DEPTH) AND CONCRETE CURB & GUTTER.
4. NEATLY SAW-CUT CONCRETE CURB & GUTTER (FULL DEPTH).
5. BLACK-OUT OR REMOVE WHITE PAVEMENT MARKING LINES. PAINT NEW 4" WIDE REFLECTIVE WHITE LINES, SEE SITE RESTORATION SHT. C3.
6. NEATLY SAW-CUT AND REMOVE (FULL DEPTH) CONC./ CURB & GUTTER AND SIDEWALK.
7. EXIST. CONC. CURB & GUTTER SHALL REMAIN.
8. REMOVE EXIST. CONC. & STEEL BARRIERS.
9. EXIST. CONC. CURB AND SIDEWALK TO REMAIN. SAW-CUT AND REMOVE HALF OF THE GUTTER (FULL DEPTH) ALONG LENGTH.
10. NEW POWERED DROP BEAM BARRIER.
11. REMOVE IN-PAVMENT DENIAL BARRIER. REMOVE STEEL COVER, ALL INTERNAL PARTS, ATTACHMENTS, PIPE, TUBING, AND LINES WITHIN DENIAL BARRIER (4 TOTAL).
12. REMOVE BARRIER CONTROL BOX AND CONTENTS TO BELOW GRADE, INCLUDING HYDRAULIC PUMP & RESERVOIR, ELECTRIC MOTOR, VEGETABLE BASE HYDRAULIC FLUID, CONDUITS, WIRES, PIPING & CONCRETE. FURNISH & INSTALL NEW BOTTOMLESS POLYMER CONCRETE ASSEMBLY BOX AND SCREW-DOWN COVER (GREEN) FLUSH IN FINISH GRADE OVER CONDUITORS AND CONDUITS. RESTORE SITE TO LAWN. CONNECT TO EXISTING CONDUITS WITH NEW CONDUIT AND EXTEND TO NEW LOCATIONS.
13. REMOVE TRAFFIC LIGHTS (2). CONCRETE BASE SHALL REMAIN. CONDUITORS MAY BE REUSED IF THEIR SIZE IS SUITABLE AND THEY ARE IN GOOD CONDITION.
14. REMOVE TRAFFIC LIGHTS (2) TO BELOW GRADE INCLUDING CONDUITS, CONDUITORS, & CONCRETE STRUCTURE. RESTORE SITE TO LAWN.
15. REMOVE & REPLACE CHAINLINK FENCE (CLF), SEE SHT. T2, F1 & D2.

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MAY 2020



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XGFG 192006  
UPGRADE MAIN GATE BARRIERS

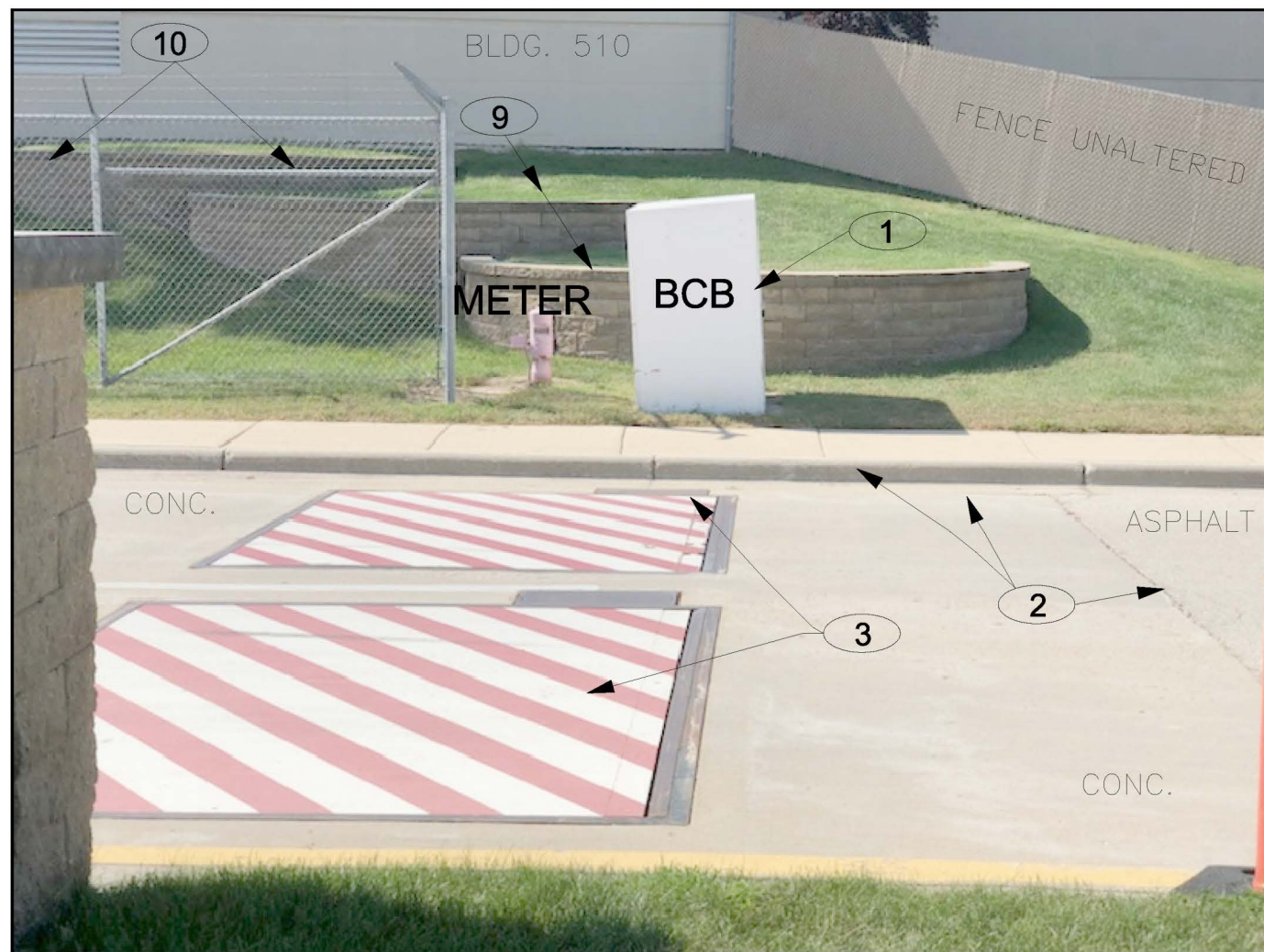
SITE REMOVAL  
PLAN - BARRIERS

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WISCONSIN AIR NATIONAL GUARD  
115<sup>TH</sup> FIGHTER WING  
TRUAX FIELD AIR NATIONAL GUARD BASE  
3110 Mitchell Street  
MADISON, WI 53704-2591  
608 245-4571

C1





1 EXIST. BARRIER 3 & 4  
EXIST. BARRIER 1 & 2, SIMILAR

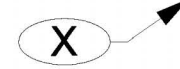


2 BARRIER CONTROL BOX (BCB)  
MOTOR LINES, RES & VALVES

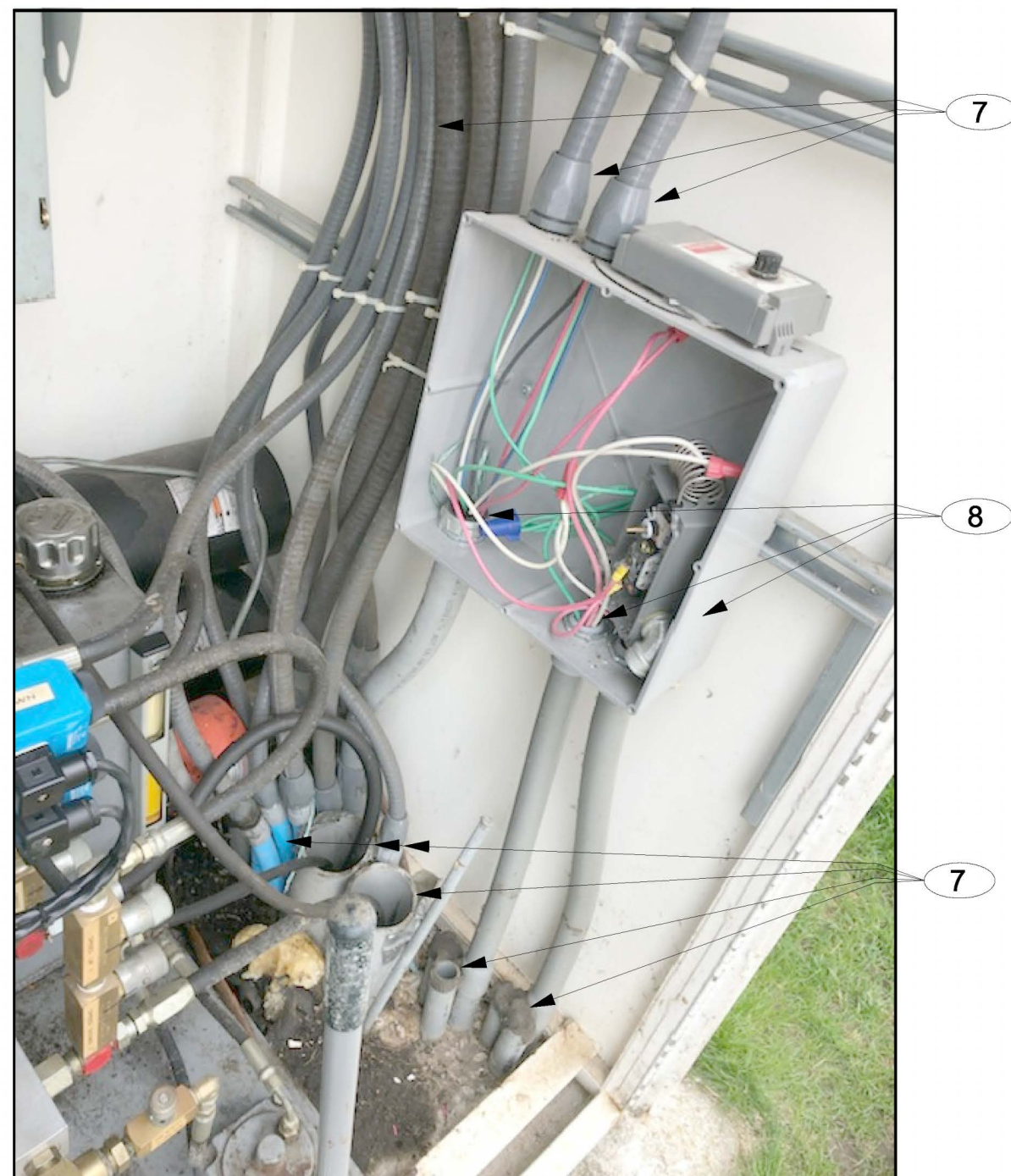


3 EXIST. BCB CONTROLS & DISCONNECT  
HYDRAULIC LINES, RES & VALVES

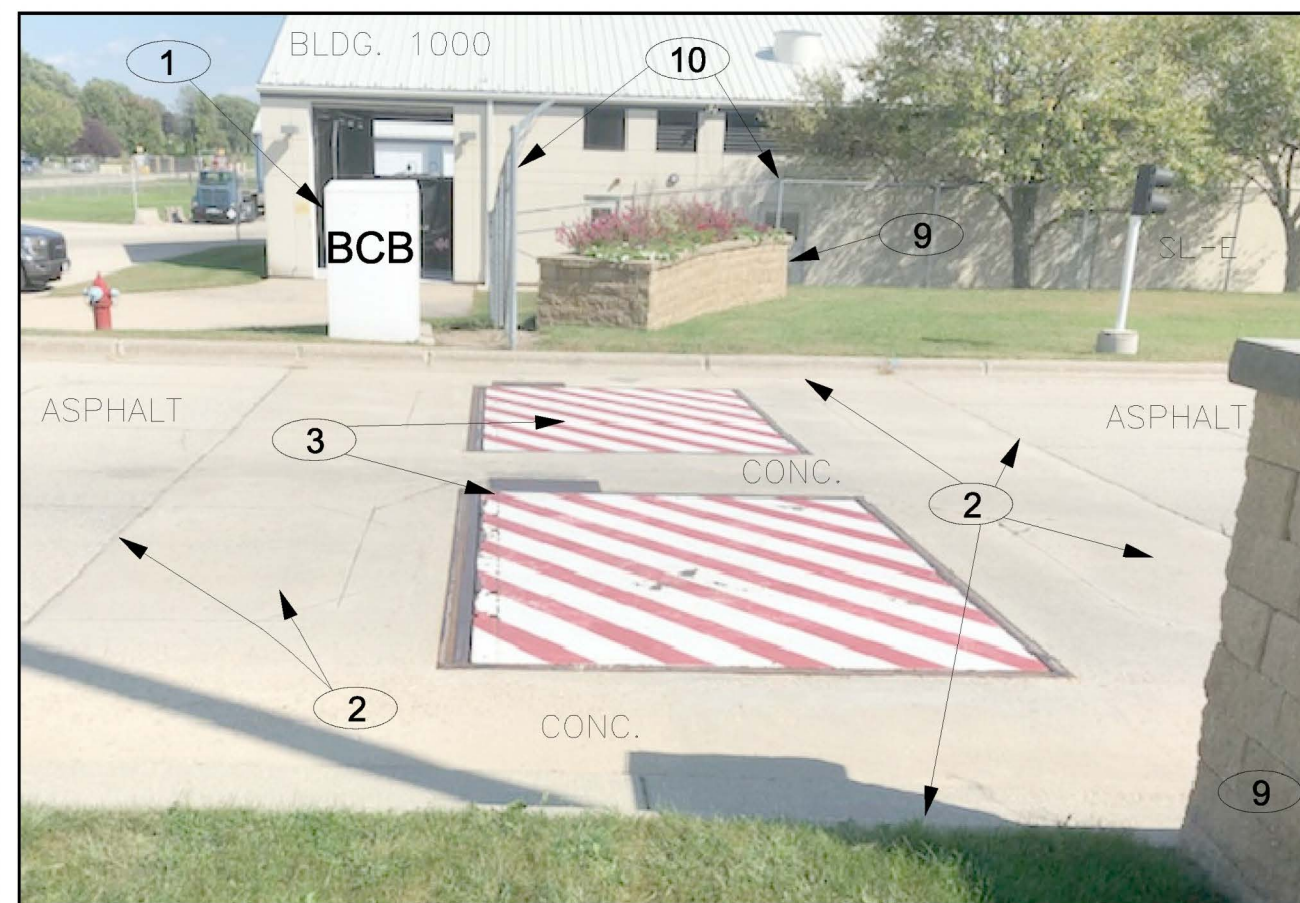
## KEYED NOTES



1. REMOVE STEEL BCB (BARRIER CONTROL BOXES). SALVAGE TO TRUAX METAL LOCATION. SEE NOTE 12 SHEET C1 & SALVAGE SITE T2.
2. REMOVE CONCRETE, CURB, GUTTER AND ASPHALT, SHT. C1.
3. REMOVE STEEL JUNCTION BOXES & STL IN-PAVEMENT BARRIERS. FILL & RESTORE AREA WITH NOT LESS-THAN 12" COMPACTED AGGREGATE #2 BASE AND 3.5" ASPHALT, SEE NOTE 11 SHEET C1 & NEW WORK SHEET C2.
4. REMOVE AND DISPOSE OF (ALL); HYDRAULIC RESERVOIR (A), FLUID, VEGETABLE OIL (B), HOSES (C), FITTINGS (D), ELECTRIC MOTOR (E) AND STEEL STRUCTURE (F).
5. REMOVE AND DISPOSE OF (ALL); LOW VOLTAGE CONTROLS (A), LINE VOLTAGE DISCONNECT (B), CONDUCTORS AND CONDUITS VIA RECYCLING METHODS.
6. REMOVE AND RECYCLE SLAVE CONTROL PANEL & CONDUCTORS.
7. CUT-OFF, PLUG AND ABANDON ALL UNUSED CONDUITS BELOW INVERT GRADE OF NEW HANDHOLE. SEE NOTE 12 SHEET C1.
8. REMOVE AND DISPOSE OF POWER JUNCTION BOX AND ALL UNUSED CONDUITS, CONDUCTORS AND CONTROLS. NEW "MASTER CONTROL PANEL" IN GUARD SHACK BLDG. 513 (NOT SHOWN) SHALL BE RECONFIGURED FOR NEW CONTROLS OF NEW BARRIERS B1, B2 & B3. NOTE B3 SHALL BE PLANNED IN THE BASE BID BUT NOT CONFIGURED UNLESS THE OPTION IS INCLUDED.
9. LANDSCAPE & WALL UNALTERED.
10. FENCE TO BE REPLACED (REFER TO SHEET T2).



4 BARRIER CONTROL BOX (BCB)



5 EXIST. BARRIER 1 & 2  
EXIST. BARRIER 3 & 4, SIMILAR



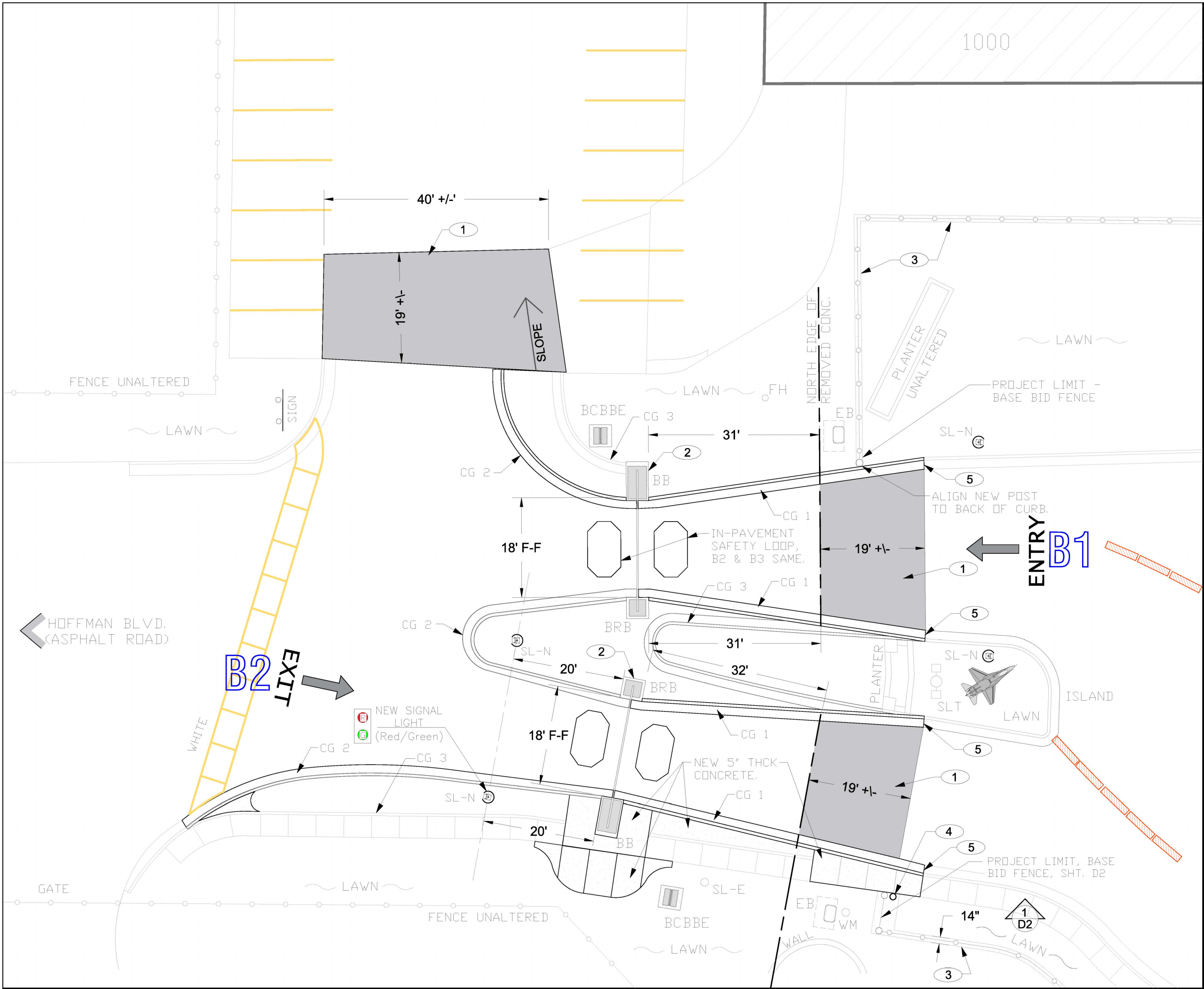
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608-245-4548 CONTRACTING OFFICER  
608-245-4571 BASE ENGINEERING  
608-245-4530 SECURITY  
608-245-4561 FIRE  
608-245-4437 SAFETY

 MAY 2020 DRAWN BY: DJW	XGFG 192006 UPGRADE MAIN GATE BARRIERS <b>BARRIER CONTROL BOX REMOVAL PLAN</b> PAGE 4 of 17
	WISCONSIN AIR NATIONAL GUARD 115 <sup>TH</sup> FIGHTER WING TRUAX FIELD AIR NATIONAL GUARD BASE 3110 Mitchell Street MADISON, WI 53704-2591 608 245-4571

**C1a**





KEYED NOTES

1. FURNISH & INSTALL NEW 3.5" THICK ASPHALT IN 2 EQUAL LIFTS OF WEARING COURSE AND FINISH COURSE OVER WELL PREPARED AGG BASE. DEPTH OF EXISTING ASPHALT IS UNKNOWN. FURNISH NEW NO. 2 AGG. BASE 10" MIN. ROUTE & SEAL NEW @ ABUTTING EXISTING PAVEMENT ALL AROUND.
2. FURNISH COMPLETE UNIT ASSEMBLY; ACTIVE ANTI-RAM RATED POWERED DROP ARM BARRIER MEETING OR EXCEEDING ASTM F2656 IMPACT CONDITION DESIGNATION M30. RSSI 3000 OR OTHER APPROVED PRODUCT. INSTALL IN ACCORDANCE WITH MFR. SPECIFICATIONS. NORMALLY IN OPEN (UP) POSITION. SEE SHT C4.
3. FURNISH & INSTALL NEW 14"W X 4" THK CONCRETE MOW STRIP CONTINUOUS UNDER NEW FENCE BOTH BASE BID AND FOR OPTIONAL FENCE RUNS WHEN OPTIONS ARE INCLUDED, SEE SHT T2.
4. FURNISH & INSTALL NEW CHAIN-LINK FENCE TERMINAL END FURNISH & INSTALL NEW 8" DIA. HOT-DIPPED GALV. STL PIPE BOLLARD W/ GROUT FILL & YELLOW PLASTIC BOLLARD COVER W/ DBL INSET REFLECTOR BANDS, BASE BID (SHT T2, & D2).
5. CONSTRUCT SMOOTH GRADUAL SLOPING TRANSITION FROM NEW CG-1 CURB & GUTTER TO ABUTTING EXISTING CG-2 CURB & GUTTER.

GENERAL NOTES

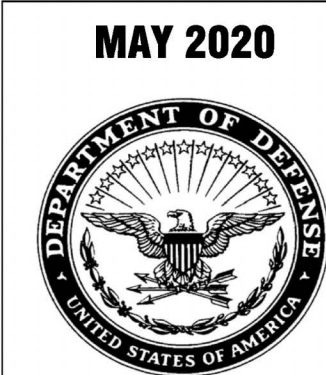
1. CONTRACTOR SHALL EMPLOY LICENSED LAND SURVEYOR TO DETERMINE HORIZONTAL AND VERTICAL CONTROL. ALL SURVEY CONTROL DATA, SITE LAYOUT MAP AND FINAL LAYOUT SHALL BE PROVIDED TO THE GOV'T FOR APPROVAL PRIOR TO NEW WORK AND "AS-BUILT" SURVEY PROVIDED WITH PROJECT CLOSEOUT.

ABBREVIATIONS:

BB: NEW BEAM BUTTRESS	EB: NEW ELECTRICAL BOX, IN-GROUND
BRB: BEAM RECEIVING BUTTRESS	E-LAWN: EXISTING LAWN
BCBBE: BEAM CONTROL & BACKUP BATTERY ENCLOSURES	SLT: STREET LIGHT POLE - EXISTING
CG 1: NEW CONC. CURB & GUTTER, DET 1	N-LAWN: NEW LAWN
CG 2: NEW CONC. CURB & GUTTER, DET 2	SL-N: SIGNAL LIGHT - NEW (4 REQ.)
CG 3: EXIST. CONC. CURB & GUTTER, DET 3	SL-E: SIGNAL LIGHT - EXISTING REMOVE NOTES 13 & 14 SHT. C1
CG 4: EXIST. CONC. CURB & GUTTER, DET 4	WM: WATER METER - EXISTING
FH: FIRE HYDRANT-EXISTING	

IMPORTANT NUMBERS

608-245-4548	CONTRACTING OFFICER
608-245-4571	BASE ENGINEERING
608-245-4530	SECURITY
608-245-4561	FIRE
608-245-4437	SAFETY



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XGFG 192006  
UPGRADE MAIN GATE BARRIERS

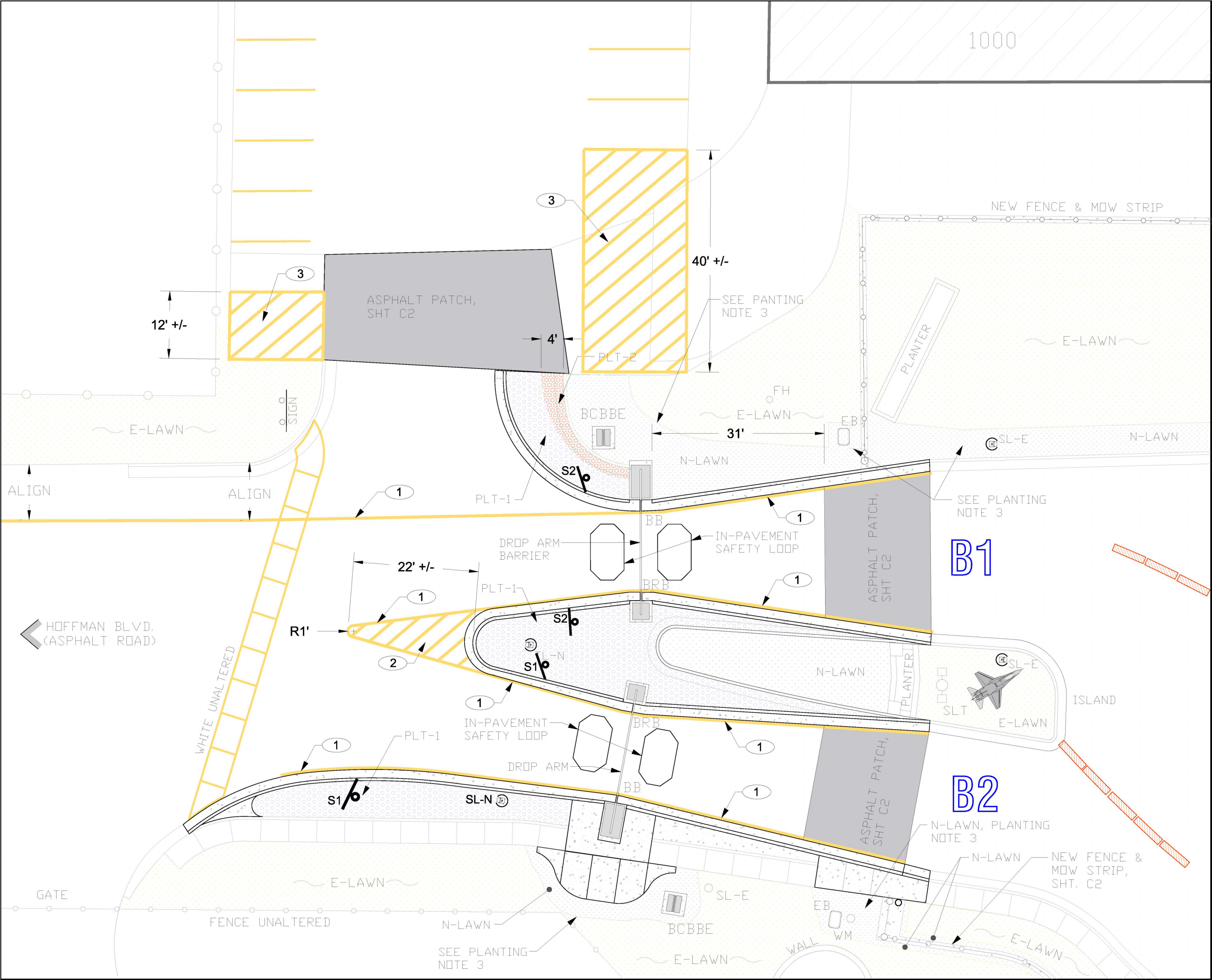
PROPOSED SITE LAYOUT

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WISCONSIN AIR NATIONAL GUARD  
115TH FIGHTER WING  
TRUAX FIELD AIR NATIONAL GUARD BASE  
3110 Mitchell Street  
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608 245-4571

C2





KEYED NOTES

- 1. FURNISH & INSTALL NEW 4" WHITE PAVEMENT MARKING.
- 2. FURNISH & INSTALL NEW 4" WHITE STRIATED PAVEMENT MARKING @ 45° - 3' SPACING.
- 3. FURNISH & INSTALL NEW 4" YELLOW STRIATED (@45° - 4' SPACING) BOX PAVEMENT MARKING.

PLANTING NOTES

- 1. N-LAWN: NEW LAWN RESTORATION. FURNISH AND INSTALL TOPSOIL NOT LESS THAN 4" DEPTH AND IN QUANTITY AS REQUIRED TO FILL AND RESTORE DISTURBED AREAS. BLEND INTO GRADE OF EXISTING LAWN. BROADCAST SPREAD NEW GRASS SEED. THATCH AND AERATE EXISTING ADJACENT AREA & BLEND IN OVER-SEED. COVER ENTIRE AREA WITH MULCH MAT. ALL NEW GRASS SEED SHALL BE PEARL'S PREMIUM (WWW.PEARLSPREMIUM.COM) FOR FULL SUN, OR OTHER PRODUCT APPROVED BY GOVERNMENT. CONTRACTOR SHALL PROVIDE SUFFICIENT HOSES, SPRINKLERS & TIMERS TO KEEP SEEDED AREAS WELL MOISTENED UNTIL HEALTHY GROWTH & MOW LAWN MINIMUM ONE TIME TO 3" HEIGHT.
- 2. E-LAWN: NO ADDITIONAL WORK REQUIRED EXCEPT AS DISTURBED BY CONTRACTOR'S OPERATION. ALL LAWN AREAS THAT ARE DISTURBED SHALL BE TREATED AS N-LAWN ABOVE.
- 3. LINE DEPICTS AVERAGE AREA AND IS NOT INTENDED AS A SPECIFIC DEFINITIVE EDGE. ALL N-LAWN AREAS ARE NON-SPECIFIC AND DEPEND UPON CONTRACTORS OPERATION IN PERFORMING THE WORK WITH REGARD TO SITE RESTORATION. CONTRACTOR SHALL BLEND N-LAWN INTO E-LAWN.
- 4. PLT-1: FURNISH POTTED "MOSS PLOX." A MIX OF COLORS IS OKAY BUT PREDOMINANT COLOR SHALL BE PURPLE WITH PINK OR WHITE MIXED RANDOMLY. GROUND COVER PLANTS SPACED NOT MORE THAN 15" APART IN EACH DIRECTION. COMPLY WITH NURSERY INSTRUCTIONS FOR PLANTING AND CARE UNTIL ESTABLISHED GROWTH. WATER AND FURNISH & INSTALL NOT-LESS-THAN, 2" THK BED OF RED SHREDDED BARK MULCH THROUGHOUT.
- 5. PLT-2: FURNISH POTTED "CHEDDAR PINK" GROUND COVER PLANTS SPACED NOT MORE THAN 12" APART IN EACH DIRECTION. COMPLY WITH NURSERY INSTRUCTIONS FOR PLANTING AND CARE UNTIL ESTABLISHED GROWTH. WATER AND FURNISH & INSTALL, NOT LESS THAN, 2" RED SHREDDED BARK MULCH SAME AS PLT-1.

ABBREVIATIONS:

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BRB: BEAM RECEIVING BUTTRESS	EB: ELECTRICAL BOX
BCBBE: BEAM CONTROL & BACKUP BATTERY ENCLOSURE	SLT: LIGHT POLE - EXISTING
CG 1: NEW CONC. CURB & GUTTER, DET 1	N-LAWN: NEW LAWN
CG 2: NEW CONC. CURB & GUTTER, DET 2	S1: "EXIT" SIGN
CG 3: EXIST. CONC. CURB & GUTTER, DET 3	S2: "DO NOT ENTER" SIGN
CG 4: EXIST. CONC. CURB & GUTTER, DET 4	SL-N: SIGNAL LIGHT - NEW (4 REQ.)
FH: FIRE HYDRANT-EXISTING	SL-E: SIGNAL LIGHT - EXISTING
	WM: WATER METER - EXISTING



S1 (DIRECTIONAL)

24" X 24" STANDARD BLACK & WHITE RELECTORIZED SIGN, TYP.



S2

24" X 24" STANDARD RED & WHITE RELECTORIZED SIGN, TYP.

NOTE: FURNISH & INSTALL SIGNS ON GREEN CHANNEL STL SIGN POST, TYPICAL.

IMPORTANT NUMBERS

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608-245-4437	SAFETY



MAY 2020

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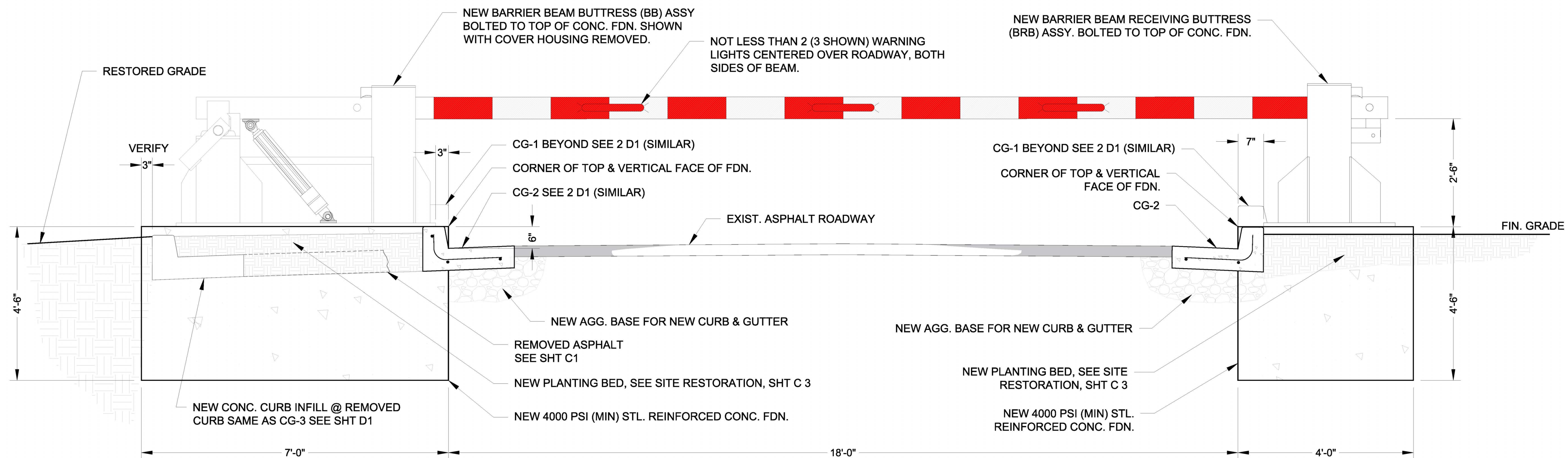
XGFG 192006  
UPGRADE MAIN GATE BARRIERS

SITE RESTORATION  
- BARRIERS

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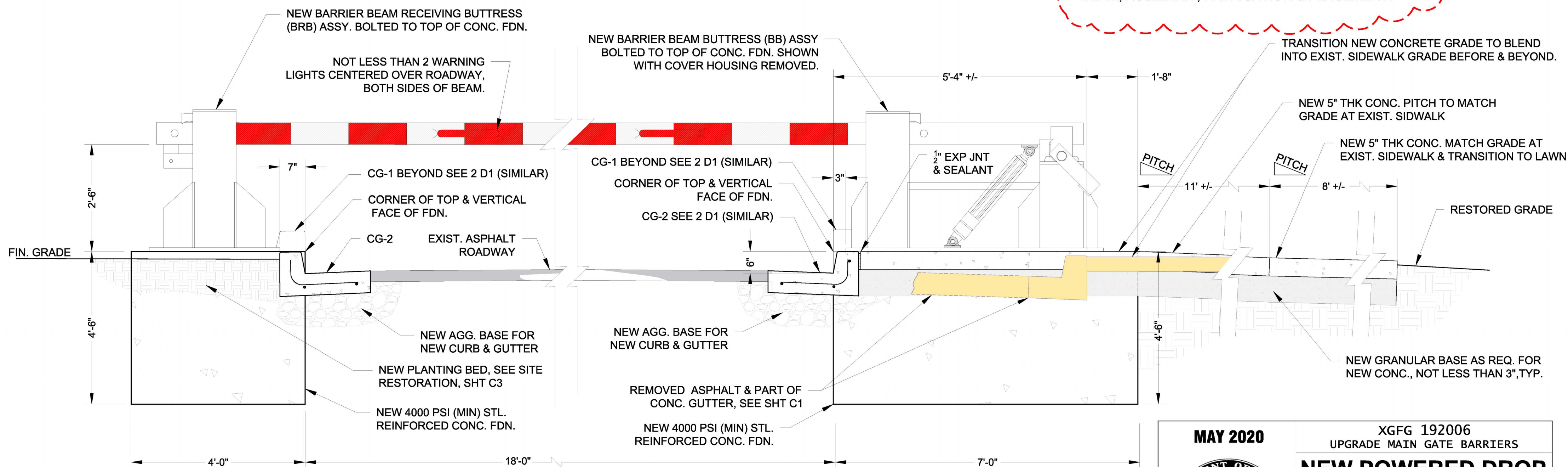
C3






**B1** 1 **POWERED DROP-BEAM BARRIER ASSY CROSS SECTION**

NOTE: DIMENSION BETWEEN FOUNDATIONS IS CRITICAL FOR EXACT MEASUREMENT OF THE BEAM. CONTRACTOR SHALL VERIFY ALL DIMENSIONS & SITE CONDITIONS FOR BEAM, ASSEMBLY, FABRICATION & PLACEMENT.

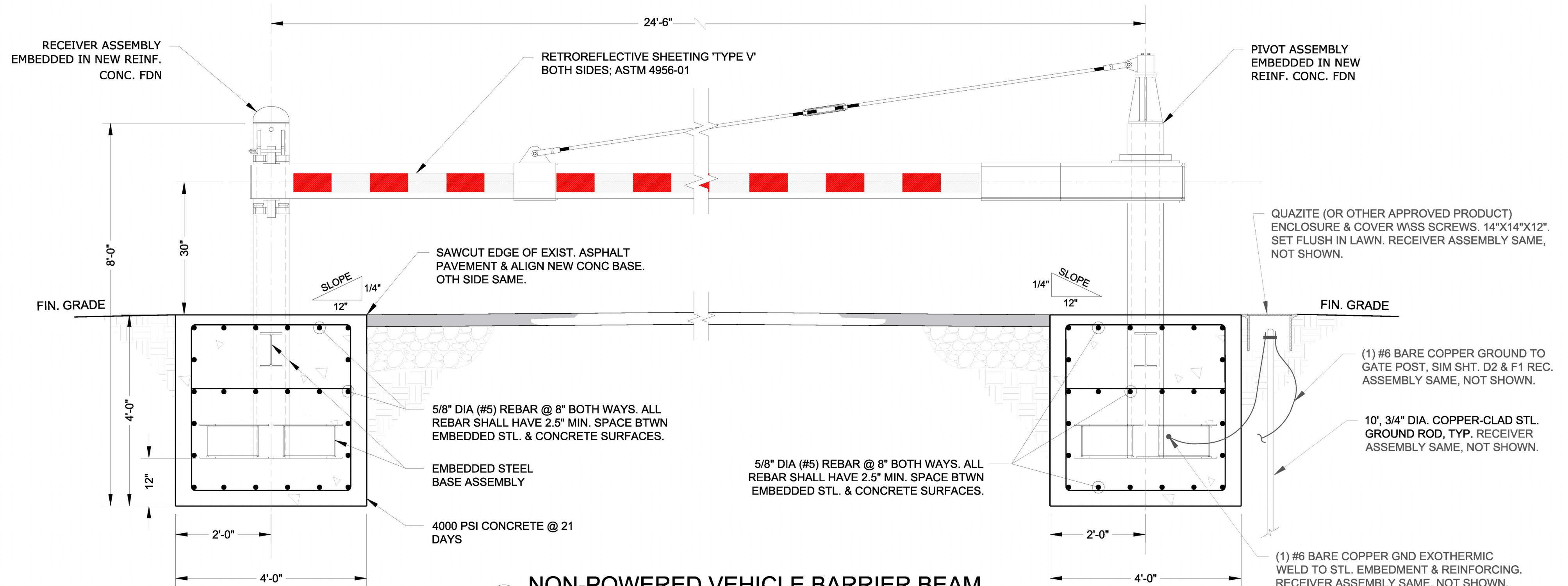


**B2** 2 **POWERED DROP-BEAM BARRIER ASSY CROSS SECTION**

 MAY 2020	XGFG 192006 UPGRADE MAIN GATE BARRIERS
	<b>NEW POWERED DROP BEAM BARRIER</b>
	PAGE 7 of 17
	WISCONSIN AIR NATIONAL GUARD 115 <sup>TH</sup> FIGHTER WING TRUAX FIELD AIR NATIONAL GUARD BASE 3110 Mitchell Street MADISON, WI 53704-2591 608 245-4571
	<b>C4</b>

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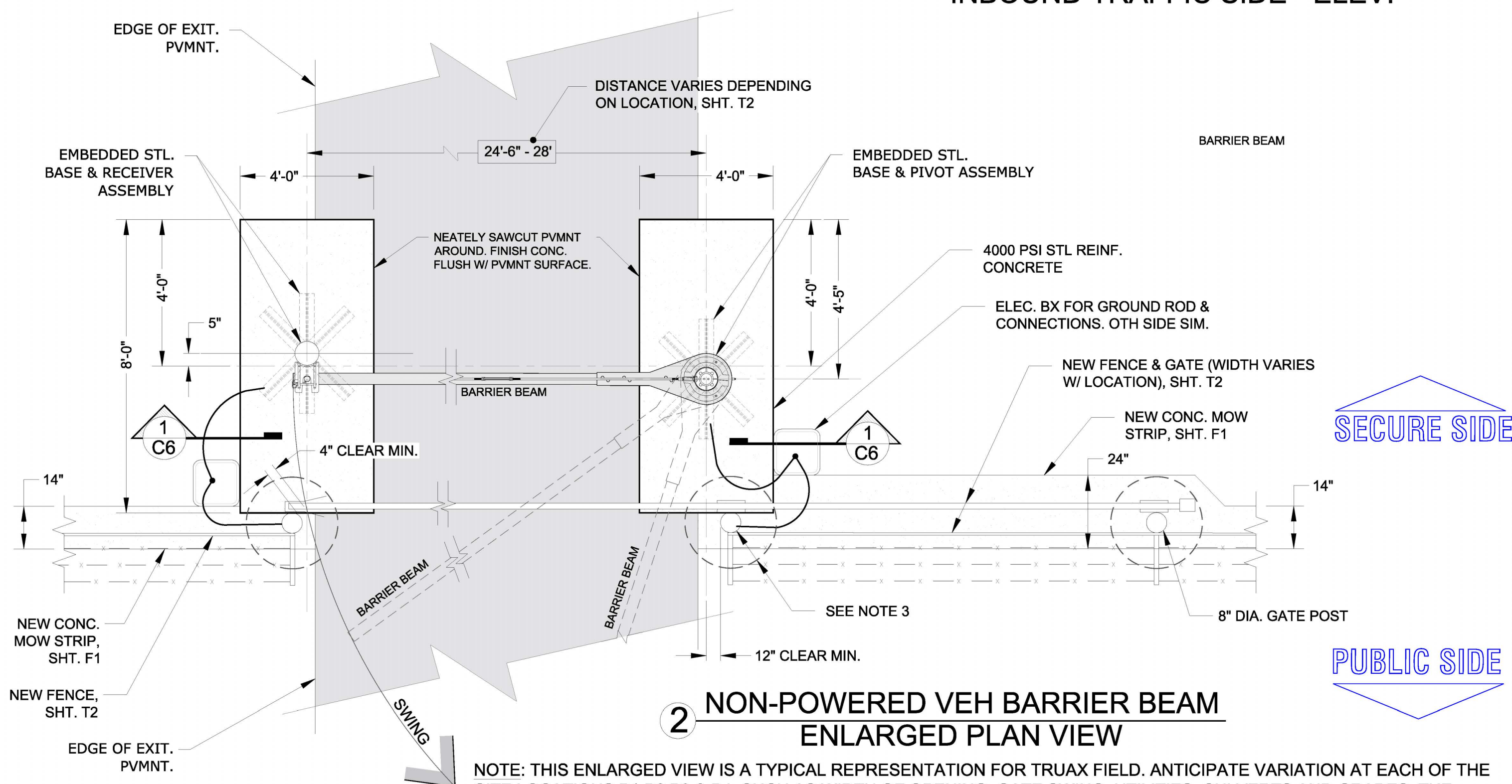




1 NON-POWERED VEHICLE BARRIER BEAM  
INBOUND TRAFFIC SIDE - ELEV.

## NOTES:

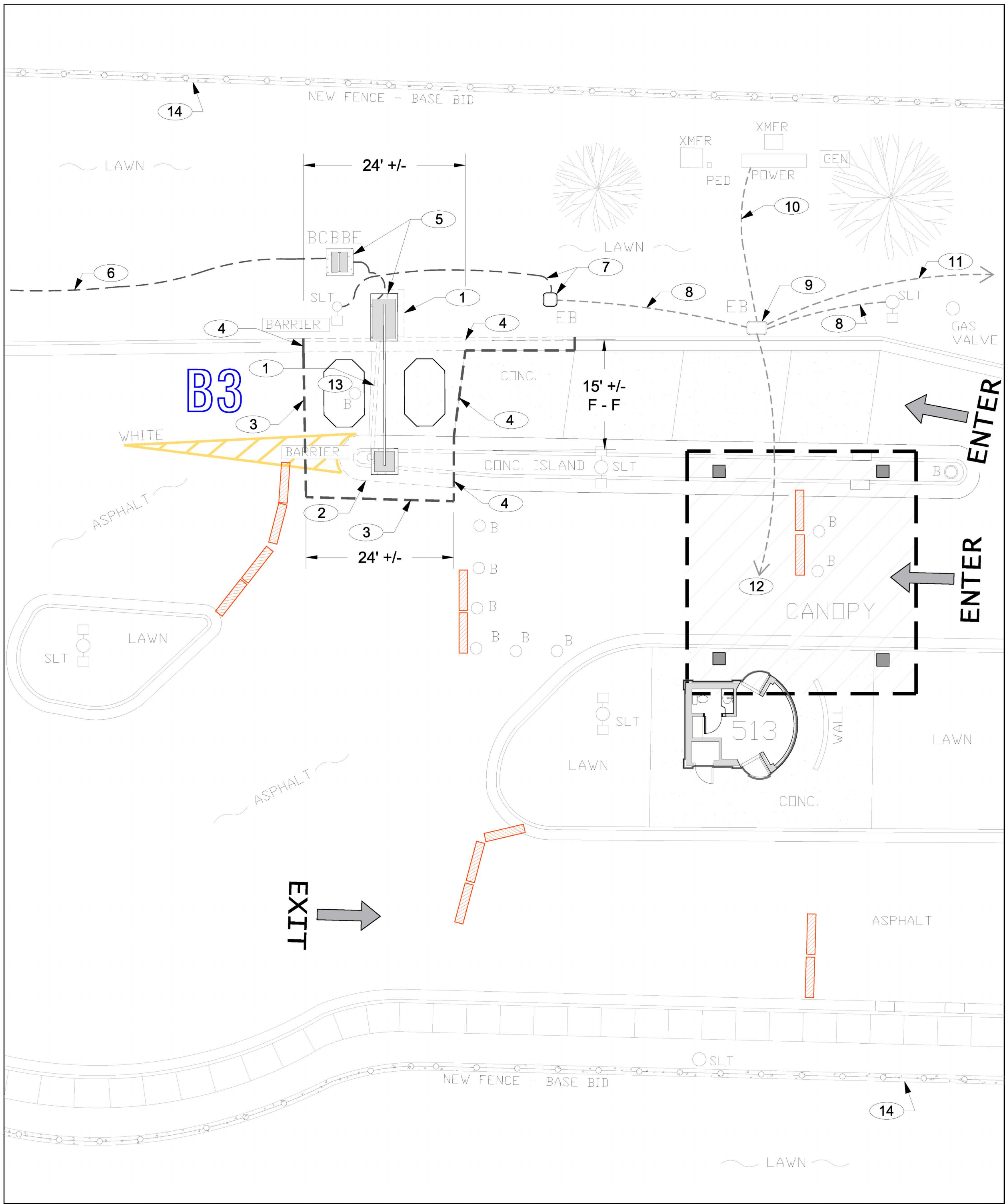
1. NEW BEAM ASSEMBLY SHOWN IS BASED ON NASATKA NMSBXII-HOR CRASH BEAM, WWW.NASATKA.COM AND IS OBSERVABLE AT OTHER LOCATIONS ON TRUAX FIELD. PROVIDE THE SAME OR SIMILAR PRODUCT AS APPROVED BY THE GOVERNMENT. COMPLY WITH MFR. REQUIREMENTS FOR INSTALLATION INCLUDING CONCRETE PLACEMENT & DESIGN. THE CONCRETE FOUNDATION DESIGN SHOWN IS THE MINIMUM REQUIRED BASED ON VEHICLE CRASH PROTECTION CAPABILITY.
2. ORIENTATION OF SWING DEPENDS ON LOCATION (SHT. T2). THE BEAM SWINGS THROUGH THE FENCE GATE OPENING & SHALL NOT BE IMPEDED BY THE GATE POSTS OR OTHER OBSTACLES & CANNOT PROTRUDE INTO TRAVELWAYS BEYOND. COORDINATE WITH FENCE INSTALLER.
3. CONSTRUCT NEW GATE POST FOUNDATION SIMILAR TO BOLLARD DETAIL (SEE SHT. D2). EXCEPTIONS: (1) WHERE THE VEHICLE BEAM OPTION IS NOT INCLUDED, CONSTRUCT 36" CYLINDRICAL FDN WITH REINFORCING BASKET AS DETAILED; (2) WHERE THE POST ABUTTS BEAM PIVOT AND RECEIVER FDN(S); POST FDN SHALL BE SEMI-CYLINDRICAL & CONNECTED TO BEAM FDN(S) W/ NOT LESS THAN 8 (FOUR PAIR) BENT #6 REBAR TIES EQUALLY SPACED VERT. COORDINATE W/FENCE INSTALLER FOR PRE-PLACEMENT OF REBAR.



NOTE: THIS ENLARGED VIEW IS A TYPICAL REPRESENTATION FOR TRUAX FIELD. ANTICIPATE VARIATION AT EACH OF THE SITE LOCATIONS B1,B2,B3 & B4, SUCH AS WIDTH OF OPENING, GATE SWING, UTILITIES, CULVERTS AND GRADES. THE CONTRACTOR SHALL INVESTIGATE, VERIFY AND MAKE ADJUSTMENTS FOR SPECIFIC REQUIREMENTS AT EACH LOCATION PRIOR TO ORDERING MATERIALS AND BEGINNING WORK.

<p>MAY 2020</p> <p>WISCONSIN AIR NATIONAL GUARD</p> <p>115<sup>TH</sup> FIGHTER WING</p> <p>TRUAX FIELD AIR NATIONAL GUARD BASE</p> <p>3110 Mitchell Street</p> <p>MADISON, WI 53704-2591</p> <p>608 245-4571</p> <p>DRAWN BY: DJW</p>	<p>XGFG 192006</p> <p>UPGRADE MAIN GATE BARRIERS</p>
	<p><b>NEW NON-POWERED VEH BARRIER BEAM</b></p>
	<p>PAGE 8 of 17</p>
	<p><b>C5</b></p>





NORTH



## KEYED NOTES

X

1. COMPLETELY REMOVE MANUAL DROP GATE, INCLUDING ALL ASSOCIATED CONCRETE AND ATTACHMENTS. SALVAGE METAL TO TRUAX METAL RECYCLING LOCATION.
2. REMOVE CONCRETE CURB, GUTTER AND CONCRETE ISLAND FOR PLACEMENT OF NEW BARRIER SYSTEM RECEIVING BUTTRESS. FULL-DEPTH. SEE SHT D1 ENLARGED PLAN.
3. SAWCUT AND REMOVE ASPHALT ROADWAY FULL DEPTH. FURNISH & PLACE NEW ASPHALT ON COMPACTED BASE SAME AS ASPHALT PATCH, SHEET C2.
4. SAWCUT CURB, GUTTER, CONCRETE ISLAND AND CONCRETE PAVEMENT. CUT PAVEMENT SECTION ALONG JOINT FULL LENGTH. FURNISH & PLACE NEW CONCRETE AS DETAILED.
5. FURNISH AND INSTALL COMPLETE UNIT ASSEMBLY; ACTIVE ANTI-RAM RATED VEHICLE POWERED DROP ARM BARRIER MEETING OR EXCEEDING ASTM F2656 IMPACT CONDITION DESIGNATION M30. RSSI 3000 OR OTHER APPROVED PRODUCT. INSTALL IN ACCORDANCE WITH MFR. SPECIFICATIONS. NORMALLY IN CLOSED (DOWN) POSITION. SIMILAR TO SHEET C1 BASE BID.
6. FURNISH AND INSTALL NEW UNDERGROUND 2" SCH 40 PVC CONDUIT W/ 6-#8 THHN HOME RUN FROM B3 & (1) 1" C W/6 CAT5E & 1-M/M FIBER. SEE SHT. E1.
7. INTERCEPT UNDERGROUND CONDUIT & CONDUCTORS. FURNISH & INSTALL NEW OPEN BOTTOM 24" X 24" X 16" QUARTZITE (OR APPROVED EQUAL) JUNCTION BOX & SCREW-DOWN COVER FLUSH IN FINISH GRADE. FURNISH & INSTALL NEW 1" PVC SCH. 40 CONDUIT AND NEW CONDUCTORS TO SLT; 2-#8, 1-#10 GND. SEE SHT. E1.
8. EXISTING UNDERGROUND SLT CONDUIT & CONDUCTORS TO REMAIN.
9. EXISTING HANDHOLE TO REMAIN.
10. EXISTING SERVICE FEEDERS TO REMAIN.
11. EXISTING CONDUIT, CONDUCTORS, BRANCH CRCT. FOR GATE MOTOR TO REMAIN.
12. EXISTING CONDUITS AND CONDUCTORS TO B513 TO REMAIN.
13. PROTECT STL BOLLARD SLEEVE AND ADJUST TO FLUSH IN NEW ASPHALT.
14. BASE BID: REMOVE CHAIN-LINK FENCE. NEATLY SALVAGE METAL TO TRUAX METAL RECYCLING LOCATION. FURNISH & INSTALL NEW CHAIN-LINK & FENCE MOW STRIP; BASE BID (SHT T2).

## IMPORTANT NUMBERS

608-245-4548 CONTRACTING OFFICER  
608-245-4571 BASE ENGINEERING  
608-245-4530 SECURITY  
608-245-4561 FIRE  
608-245-4437 SAFETY

MAY 2020



DRAWN BY: DJW

XGFG 192006  
UPGRADE MAIN GATE BARRIERS

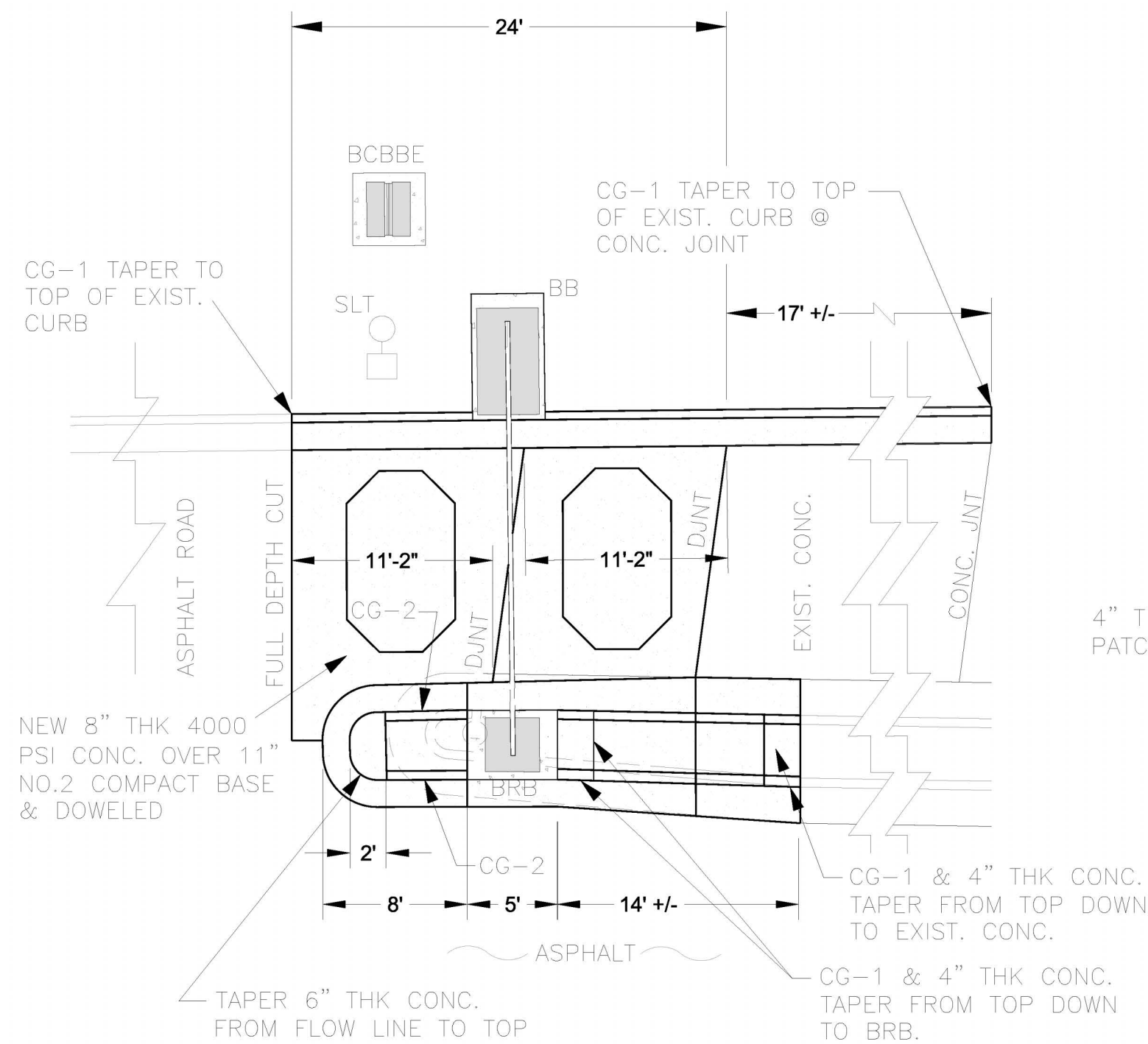
## OPTON 2 PLAN

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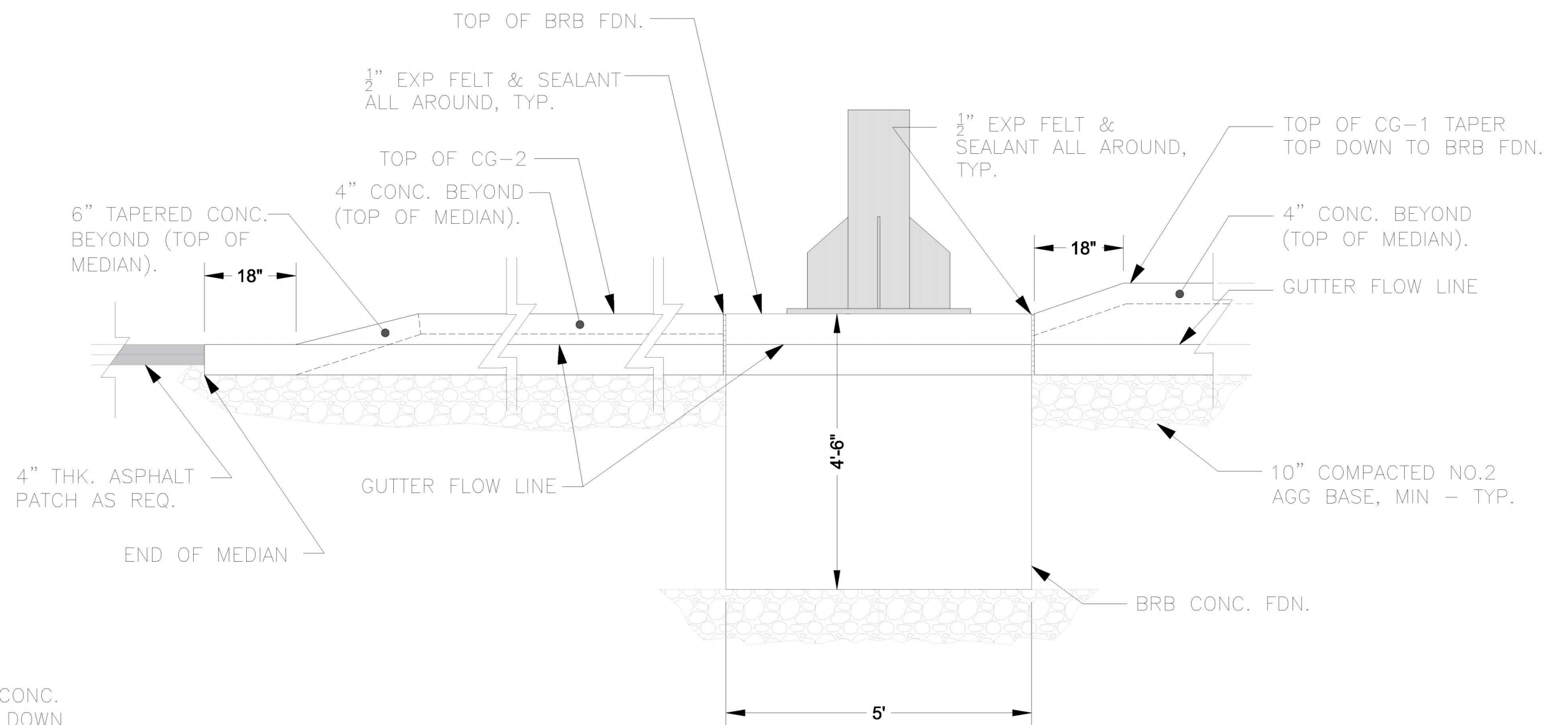
WISCONSIN AIR NATIONAL GUARD  
115<sup>TH</sup> FIGHTER WING  
TRUAX FIELD AIR NATIONAL GUARD BASE  
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608 245-4571

C6

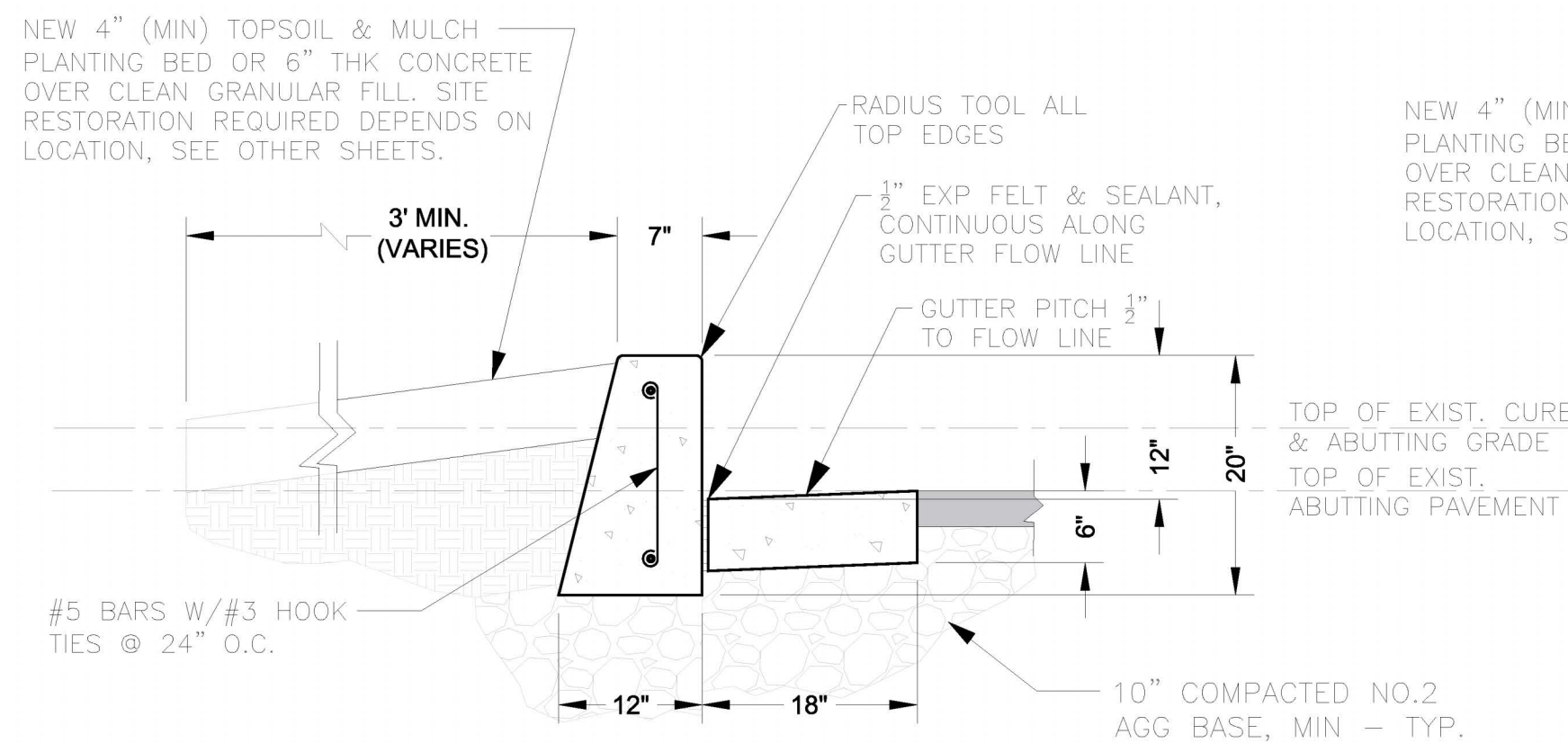




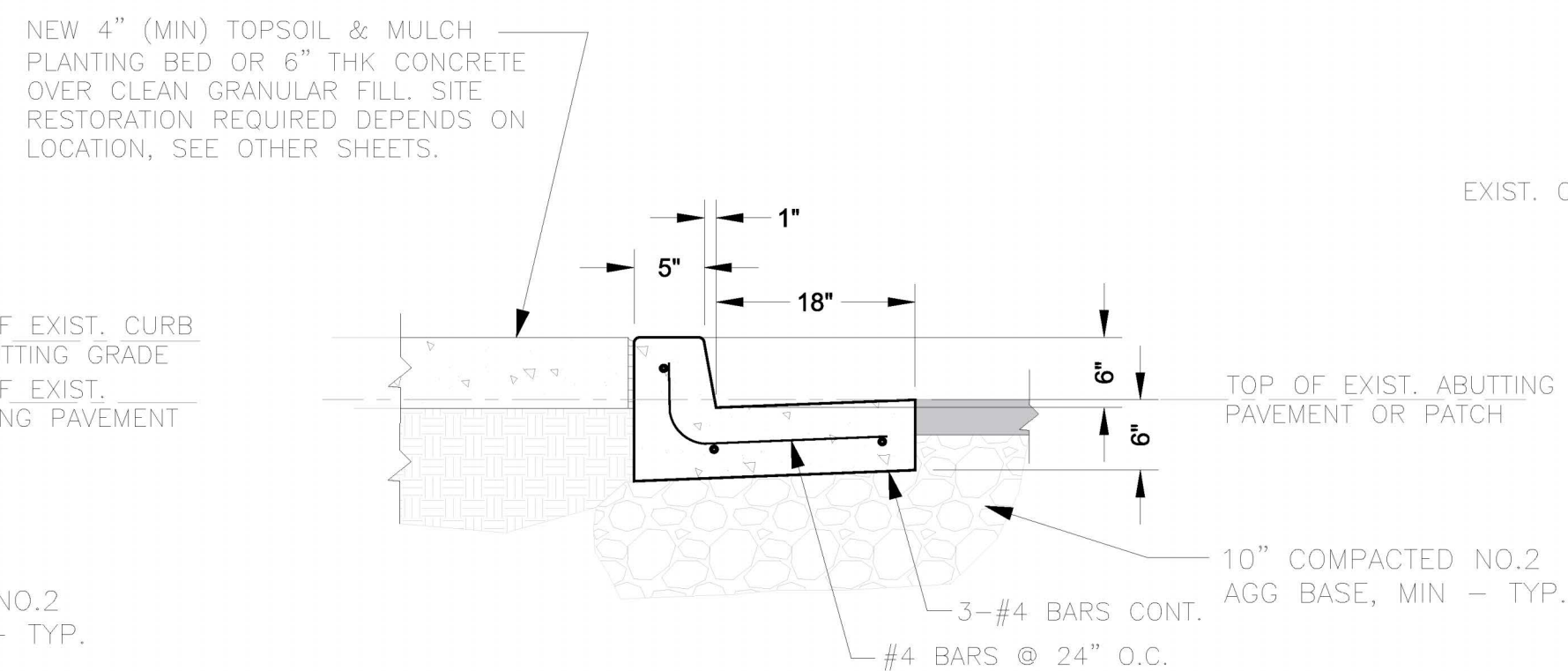
1 **OPTION 2 - ENLARGED PLAN**



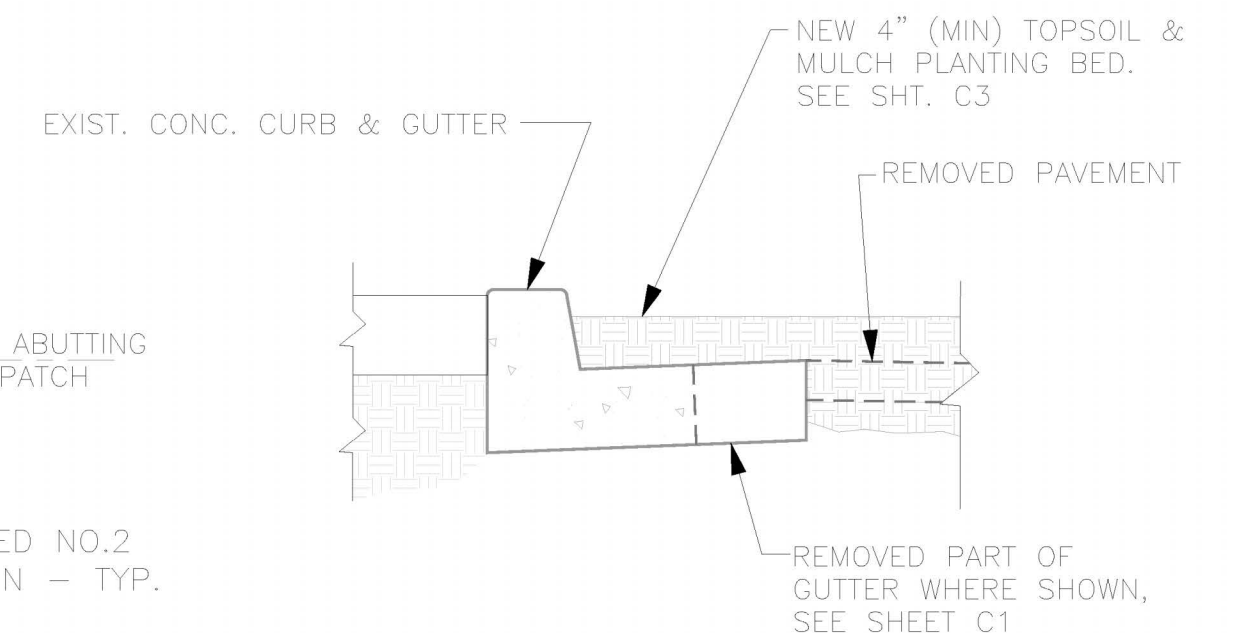
2 **OPTION 2 - RECEIVING BUTTRESS  
MEDIAN RECONSTRUCTION**



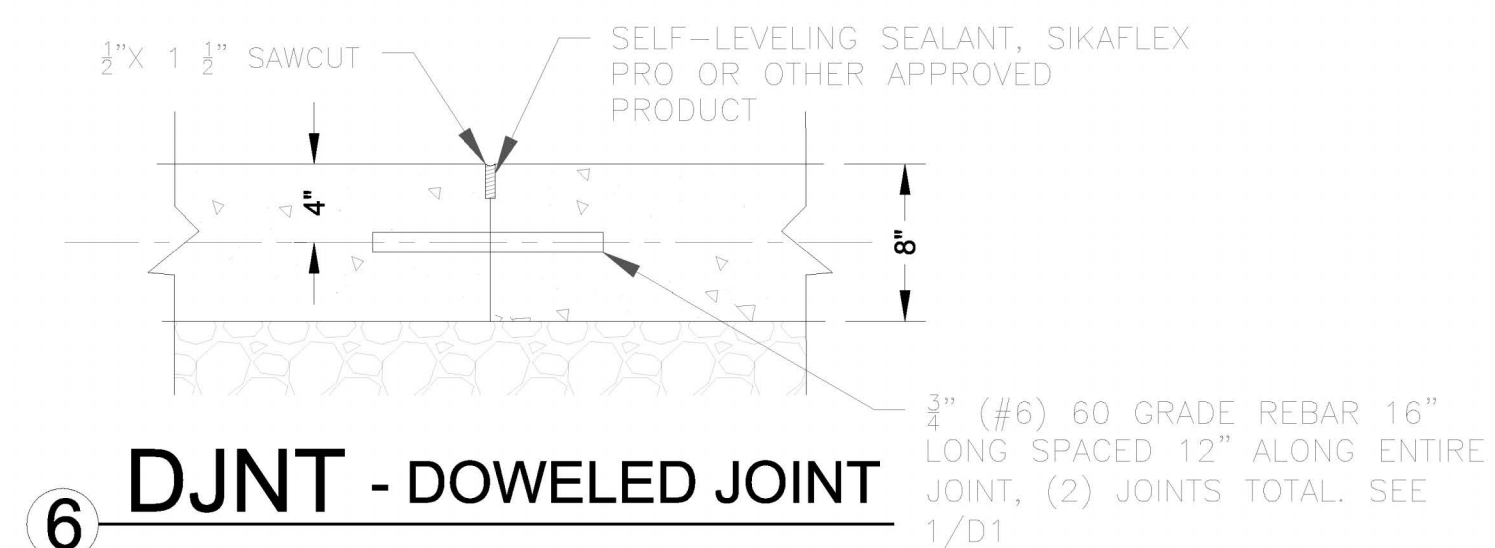
3 **CG-1 CONC. CURB & GUTTER**  
NEW CONC. 3000 PSI MIN.  
FIBER REINFORCED



4 **CG-2 CONC. CURB & GUTTER**  
NEW CONC. 3000 PSI MIN.  
FIBER REINFORCED



5 **CG-3 CONC. CURB & GUTTER**  
EXISTING



6 **DJNT - DOWELED JOINT**

### IMPORTANT NUMBERS

608-245-4548  
608-245-4571  
608-245-4530  
608-245-4561  
608-245-4437

CONTRACTING OFFICE  
BASE ENGINEERING  
SECURITY  
FIRE  
SAFETY

 MAY 2020	XGFG 192006 UPGRADE MAIN GATE BARRIERS
	<b>DETAILS</b>
	PAGE 10 of 17
	WISCONSIN AIR NATIONAL GUARD 115 <sup>TH</sup> FIGHTER WING TRUAX FIELD AIR NATIONAL GUARD BASE 3110 Mitchell Street MADISON, WI 53704-2591 608 245-4571

**D1**

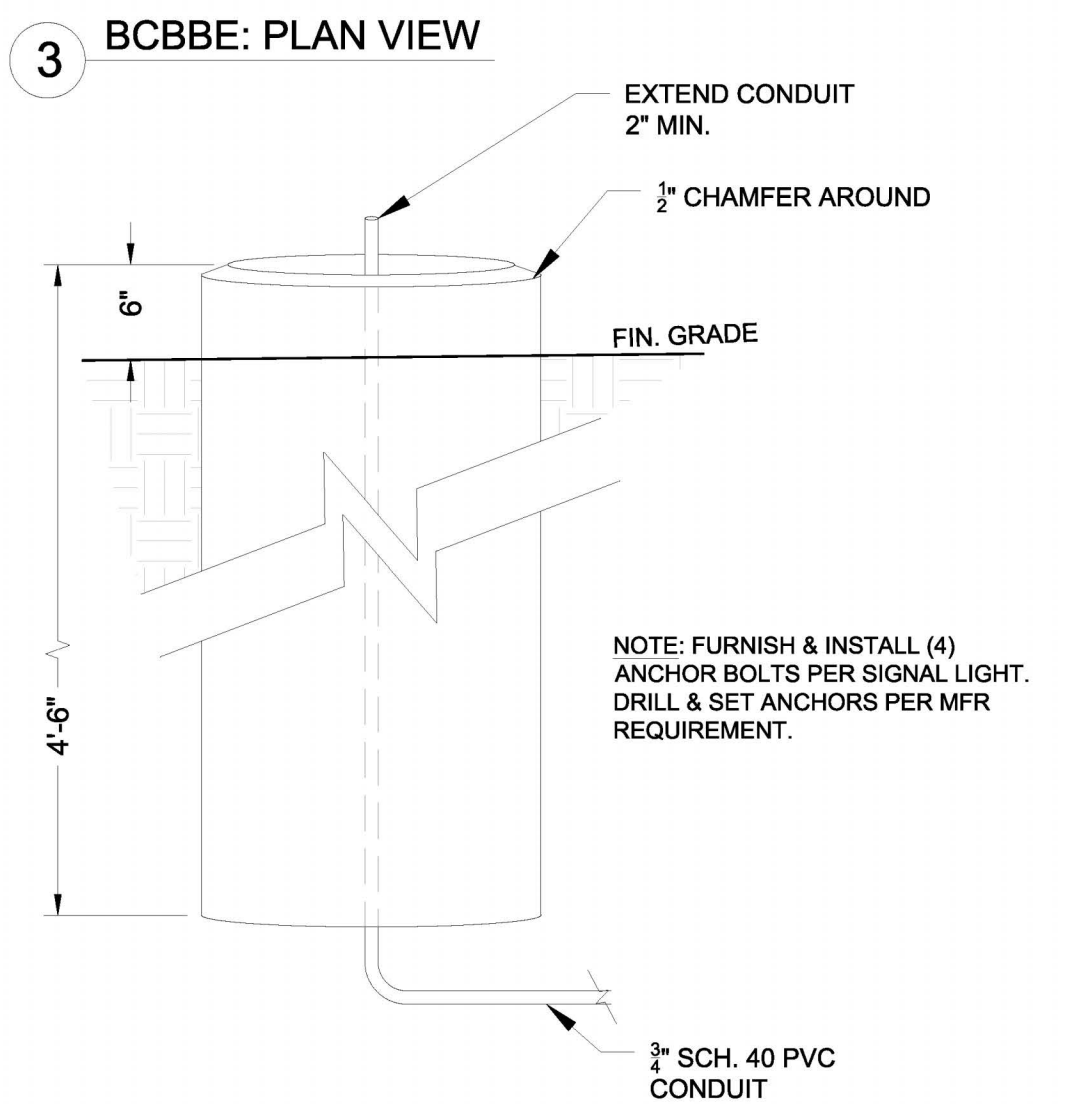
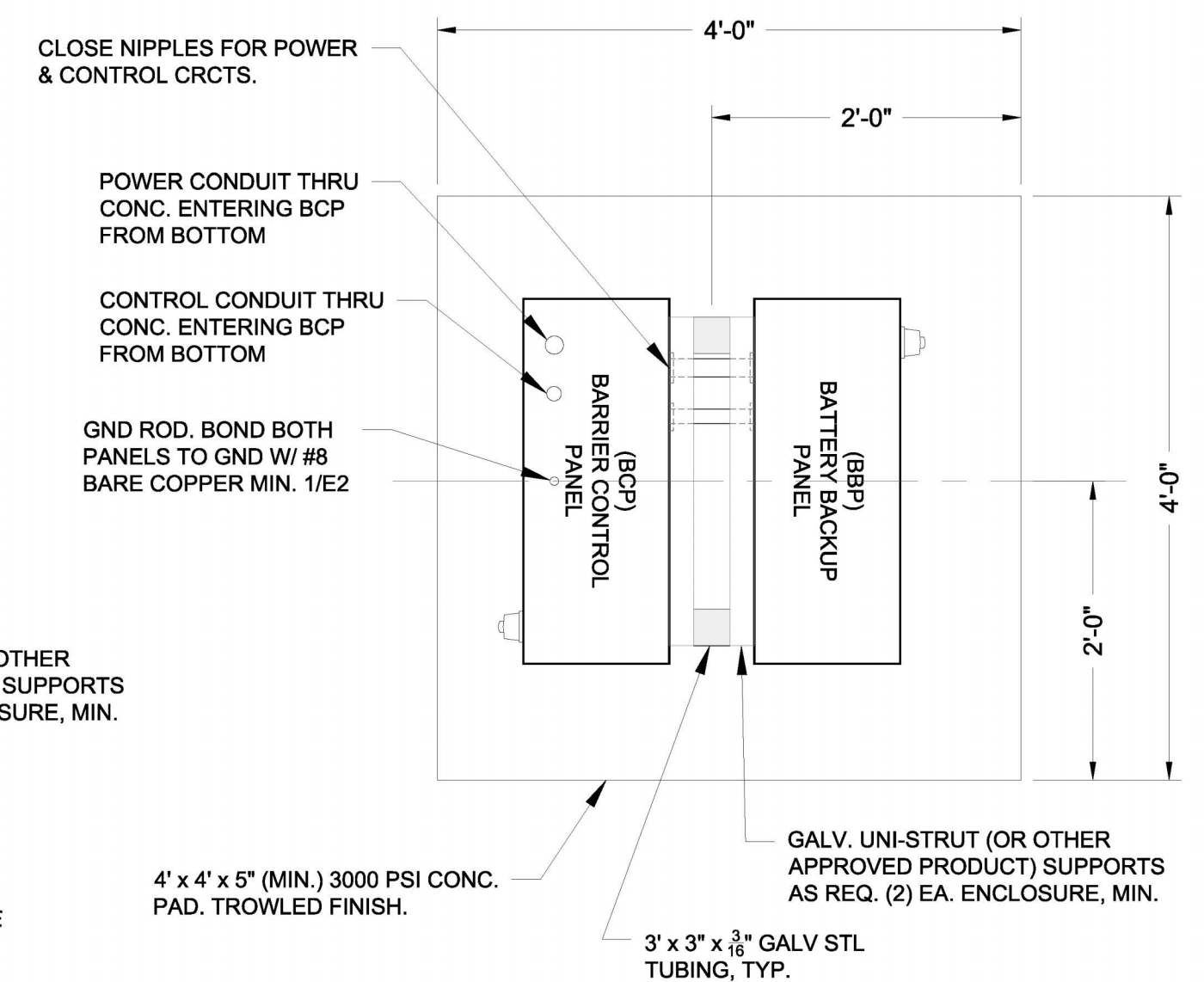
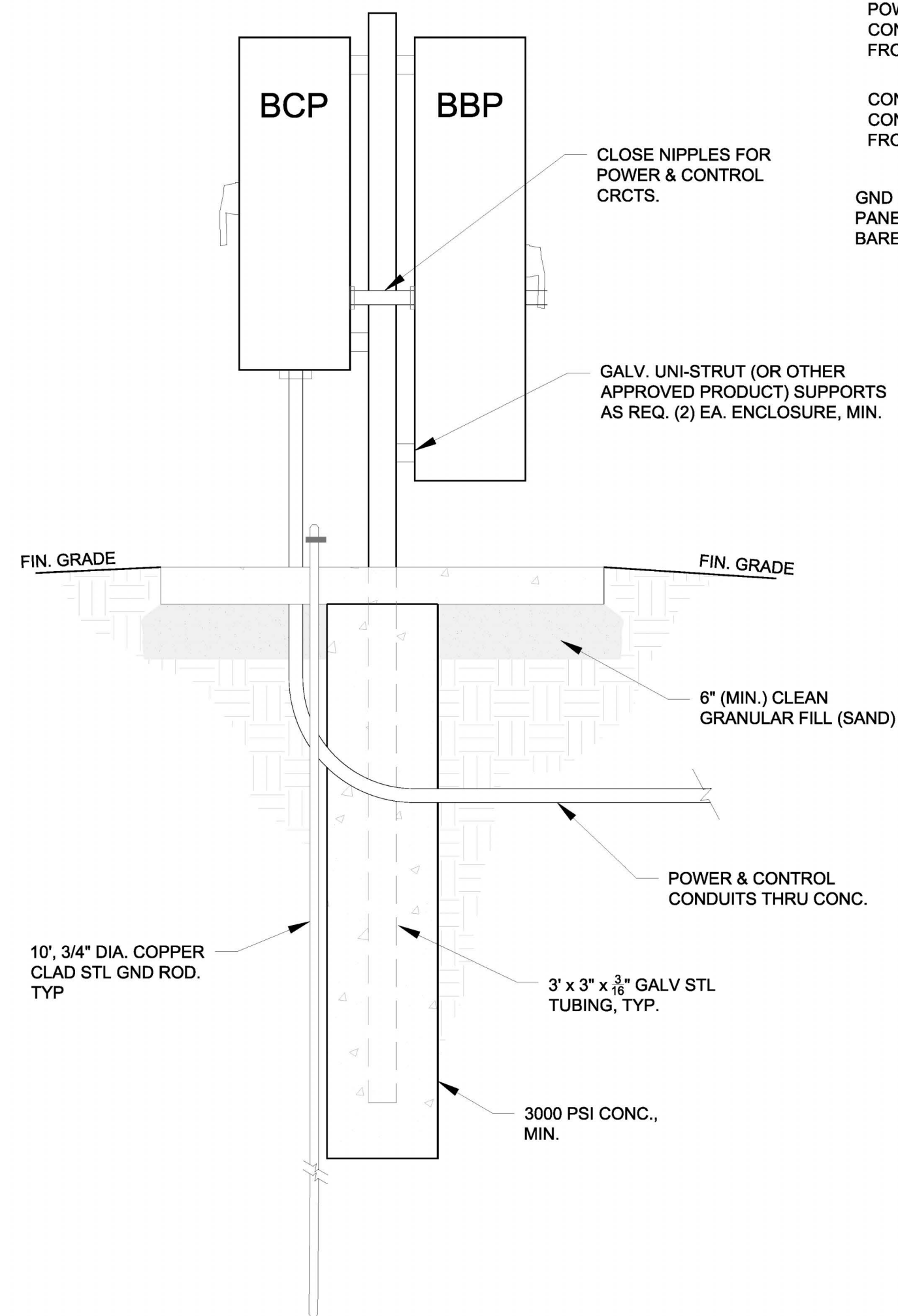
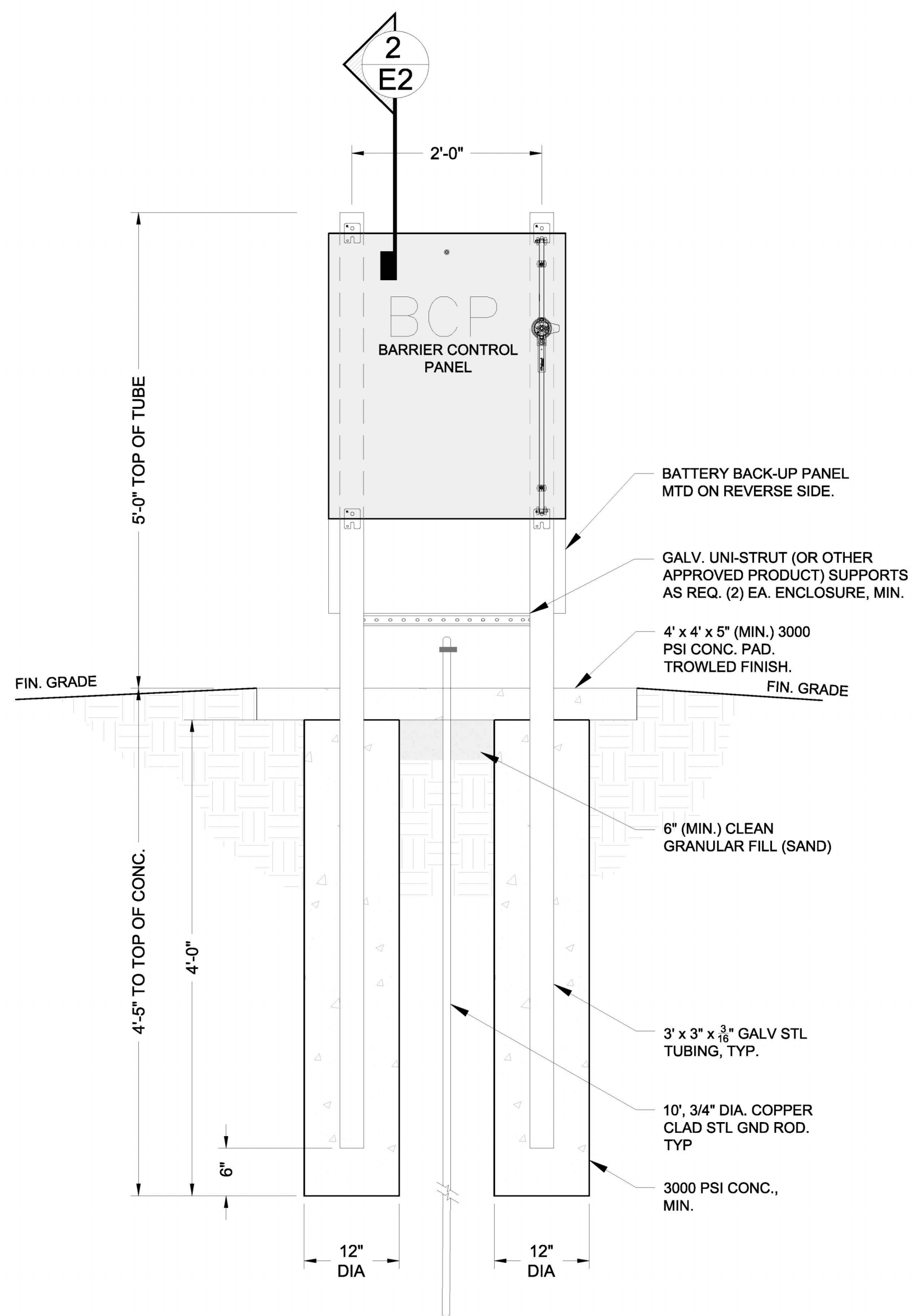












## IMPORTANT NUMBERS

**608-245-4548 CONTRACTING OFFICE**  
**608-245-4571 BASE ENGINEERING**  
**608-245-4530 SECURITY**  
**608-245-4561 FIRE**  
**608-245-4437 SAFETY**



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MAY 2020

XGFG 192006  
UPGRADE MAIN GATE BARRIERS

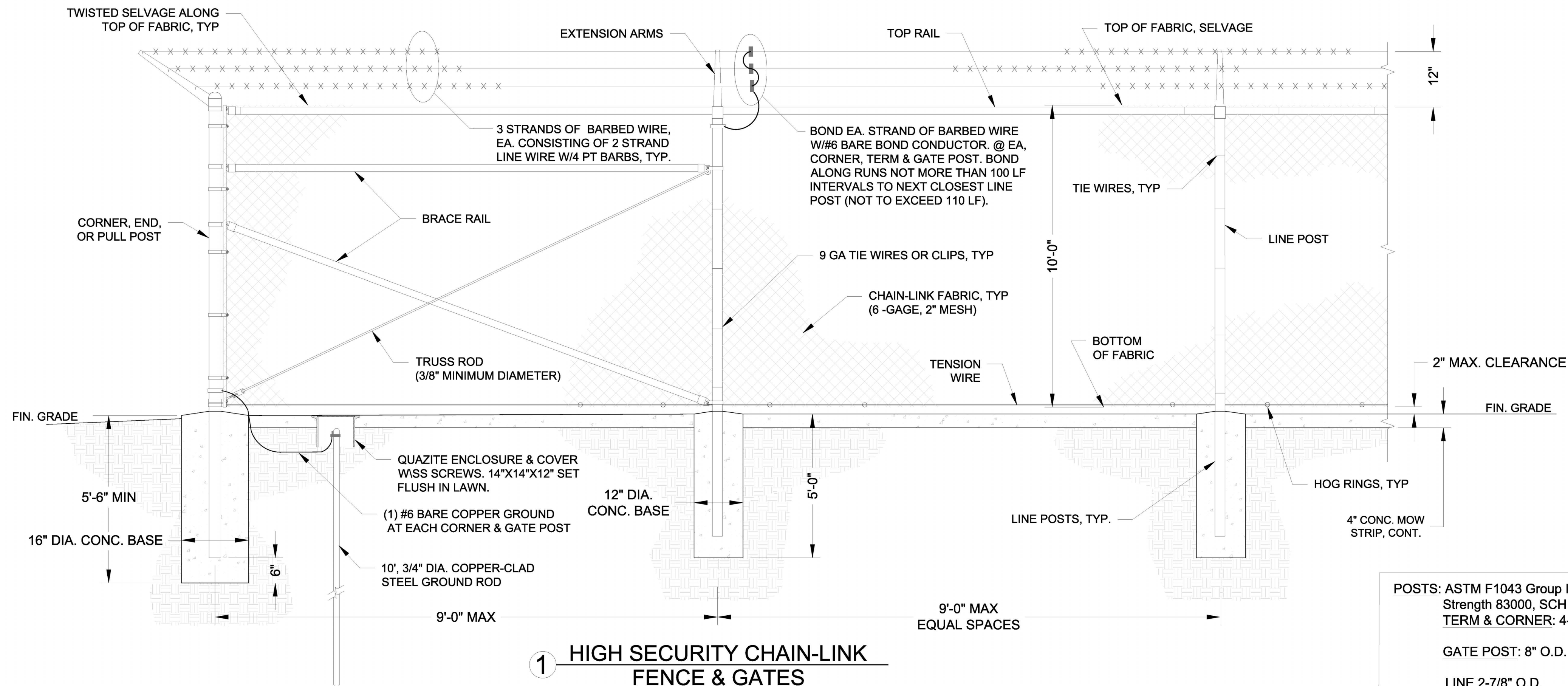
## ELECTRICAL DETAILS

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WISCONSIN AIR NATIONAL GUARD  
115<sup>TH</sup> FIGHTER WING  
TRUAX FIELD AIR NATIONAL GUARD BASE  
3110 Mitchell Street  
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608 245-4571

## E2





**POSTS:** ASTM F1043 Group IA High Strength 83000, SCH 40  
**TERM & CORNER:** 4-1/2" O.D.

**GATE POST:** 8" O.D. (C6 NOTE 3)

**LINE** 2-7/8" O.D.

**FABRIC:** 6 GA X 2" MESH

**TOP RAIL:** 1-5/8" O.D.

**MIDDLE & BRACE RAIL:** 1-5/8" O.D

**BOTTOM RAIL:** 1-5/8" O.D

**POST CAPS:** DOME STYLE

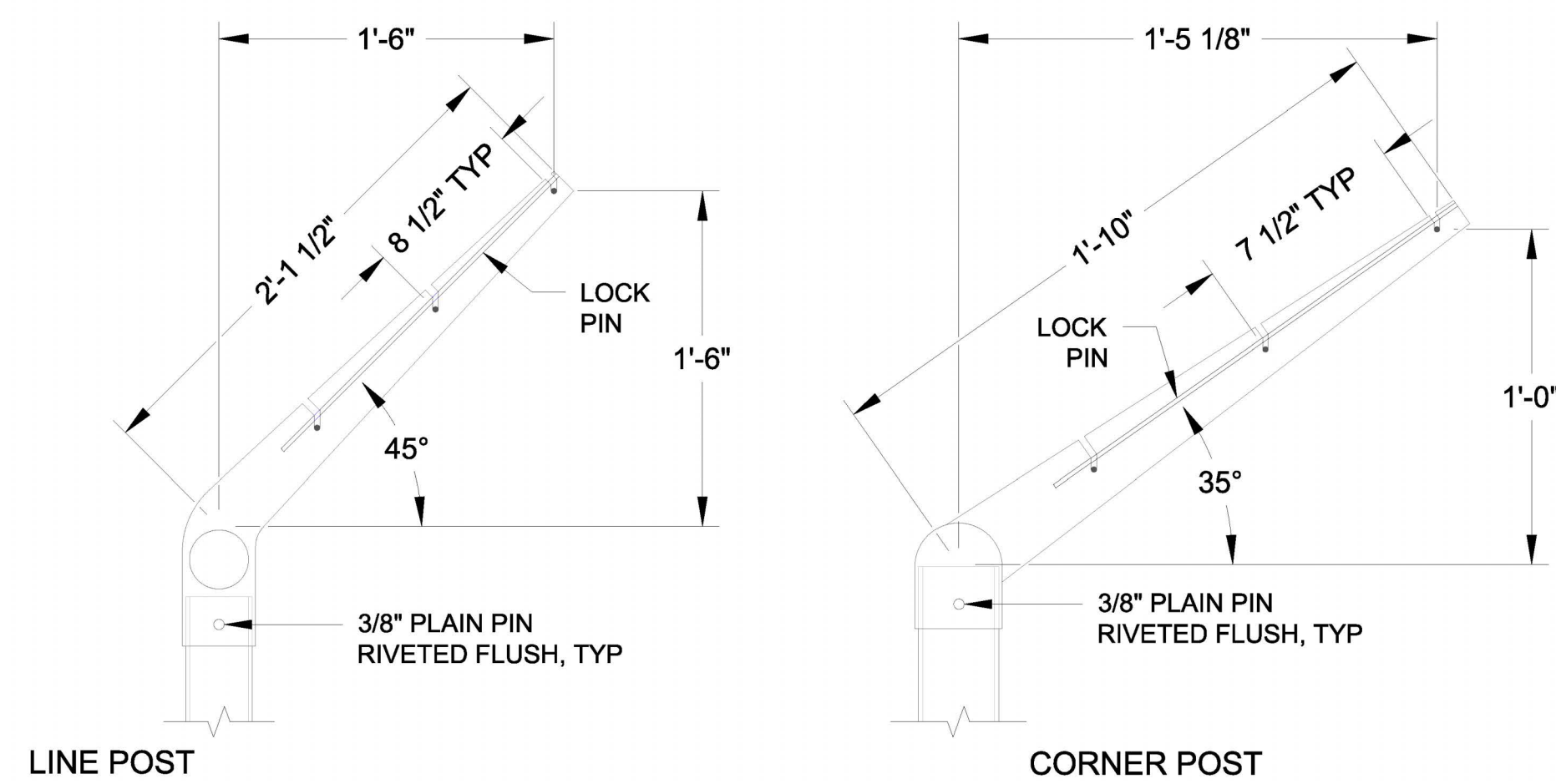
**TIES:** 9GA ALUMINUM

**TOP (BARBED)WIRES:** ASTM 121, 12 GA

**MOW STRIP:** CONTINUOUS 14" BY 4" FIBER REINFORCED.

**ALL CONCRETE:** 4000 PSI

CONTRACTOR SHALL COORDINATE EVERY DAY'S PROGRESS THROUGH TRUAX SECURITY. THERE SHALL BE NO REMOVED FENCE WITHOUT TRUAX SECURITY APPROVAL AND TEMPORARY SECURITY MEASURES IN PLACE.

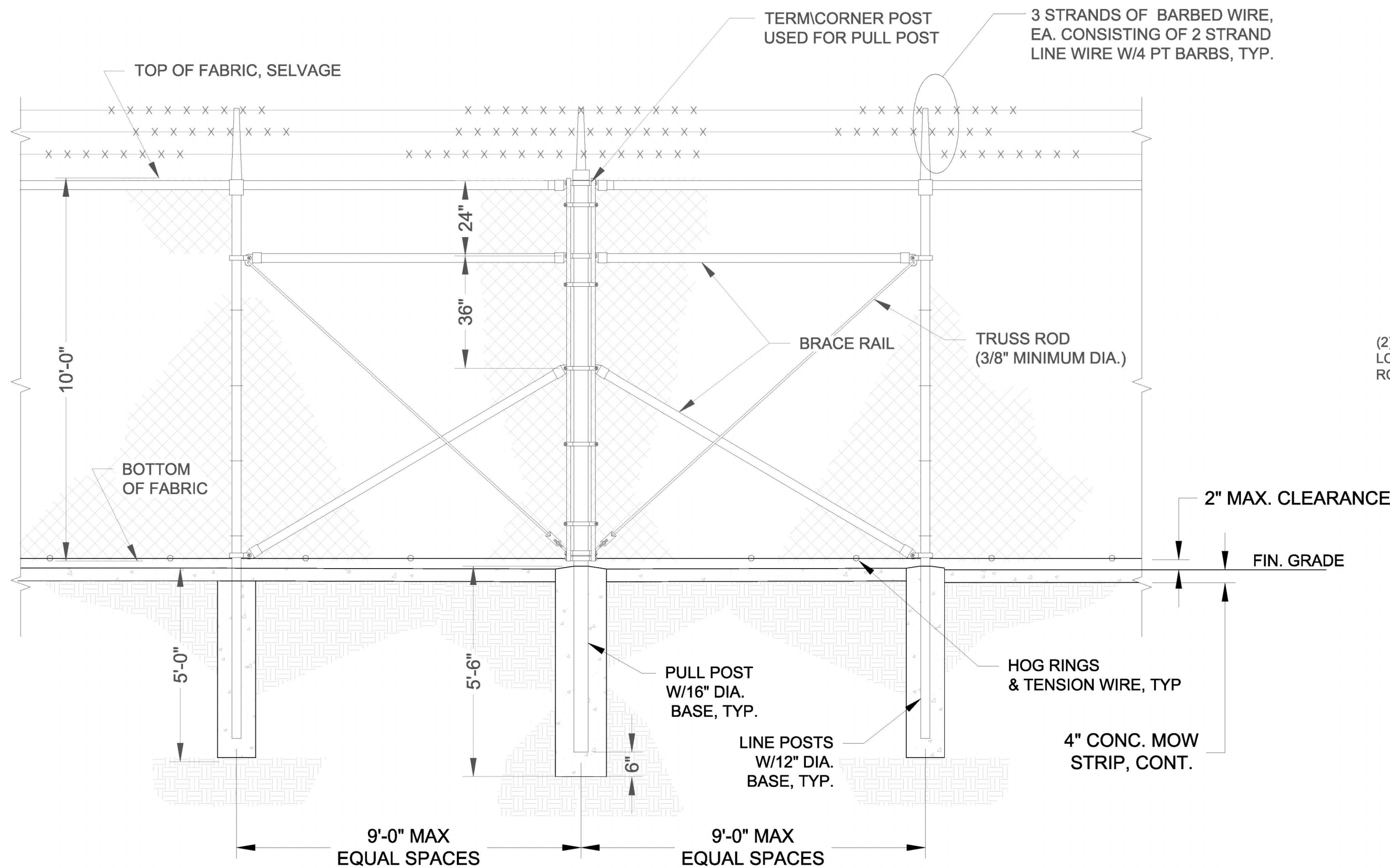


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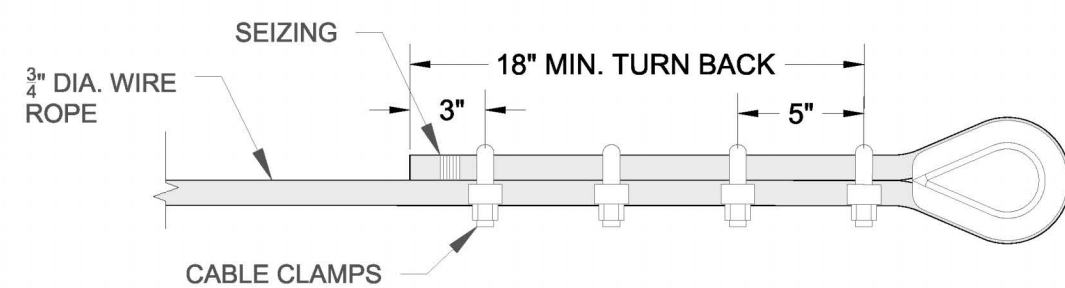
608-245-4548 CONTRACTING OFFICER  
 608-245-4571 BASE ENGINEERING  
 608-245-4530 SECURITY  
 608-245-4561 FIRE  
 608-245-4437 SAFETY

 MAY 2020 DRAWN BY: DJW	XGFG 192006 UPGRADE MAIN GATE BARRIERS
	<b>NEW HIGH-SECURITY CHAINLINK (CLF) DETAILS</b> PAGE 14 of 17
	WISCONSIN AIR NATIONAL GUARD 115 <sup>TH</sup> FIGHTER WING TRUAX FIELD AIR NATIONAL GUARD BASE 3110 Mitchell Street MADISON, WI 53704-2591 608 245-4571
	<b>F1</b>





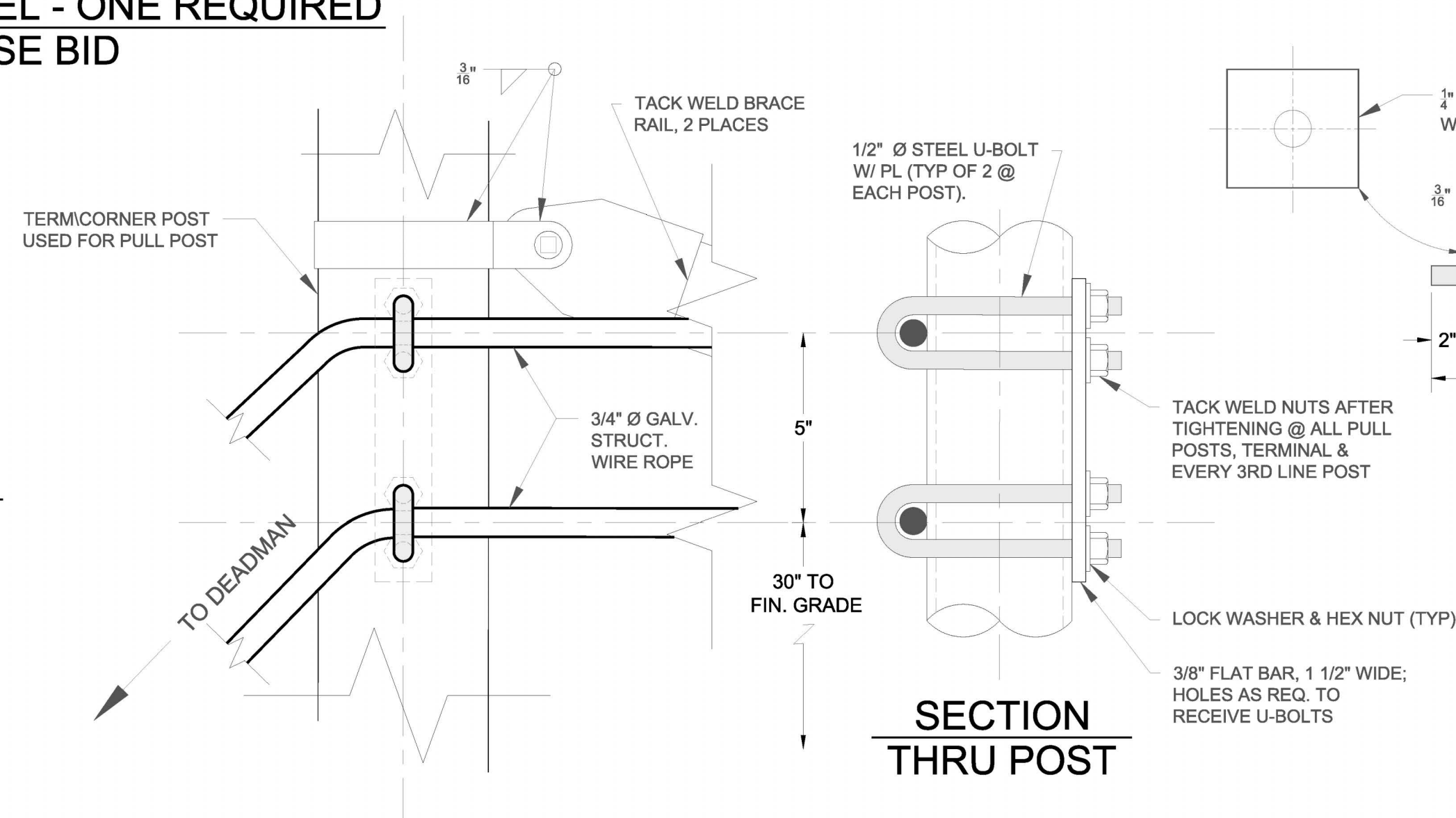
**1 CLF BRACE PANEL - ONE REQUIRED BASE BID**



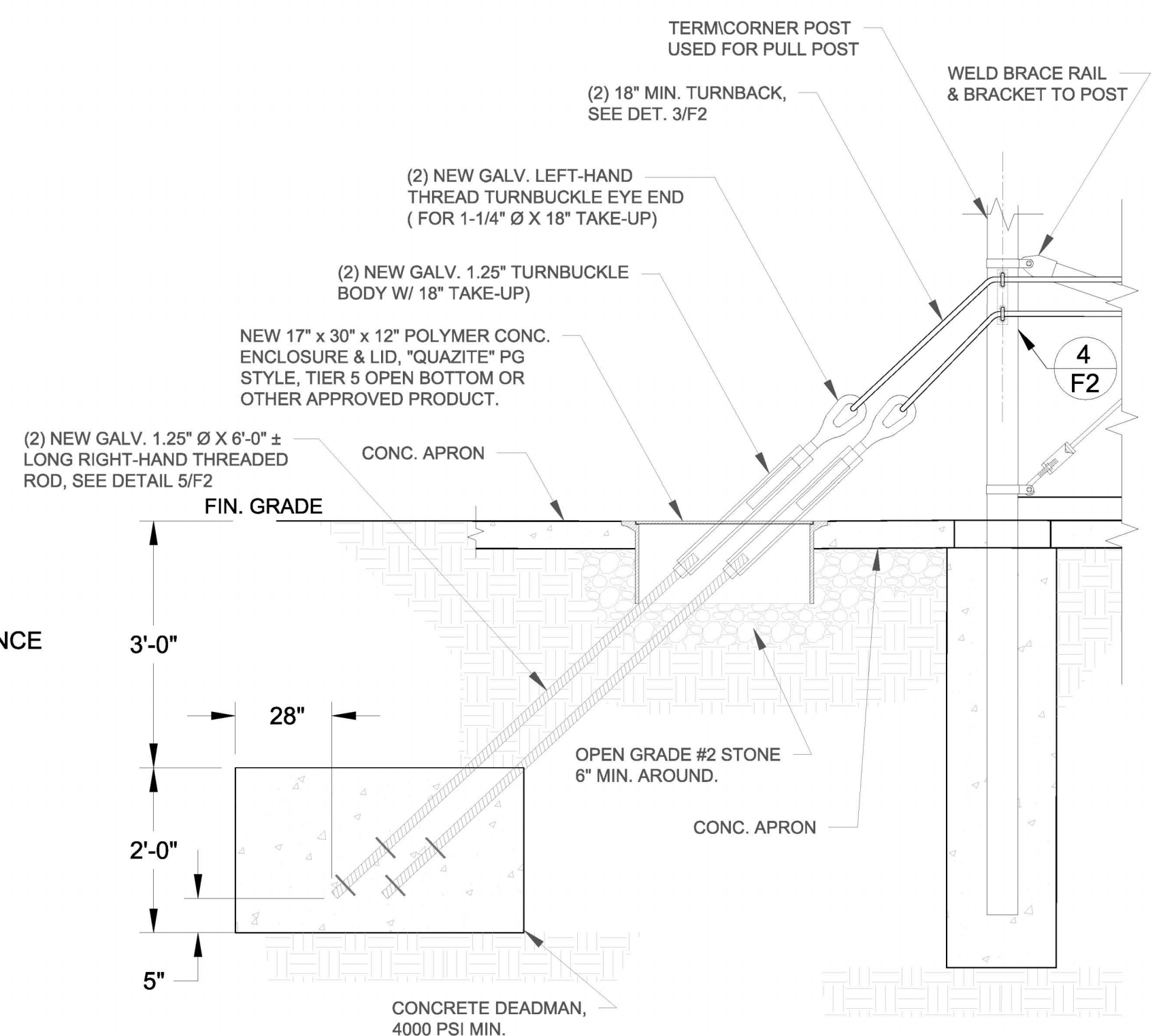
**3 TYPICAL TURNBACK AND CLAMP DETAIL**

NOTE: INSTALL SADDLE OF CLIPS ON LIVE SIDE (AS SHOWN), SEIZE CUT END OF WIRE ROPE, AND TORQUE NUTS PER MANUFACTURER'S RECOMMENDATIONS.

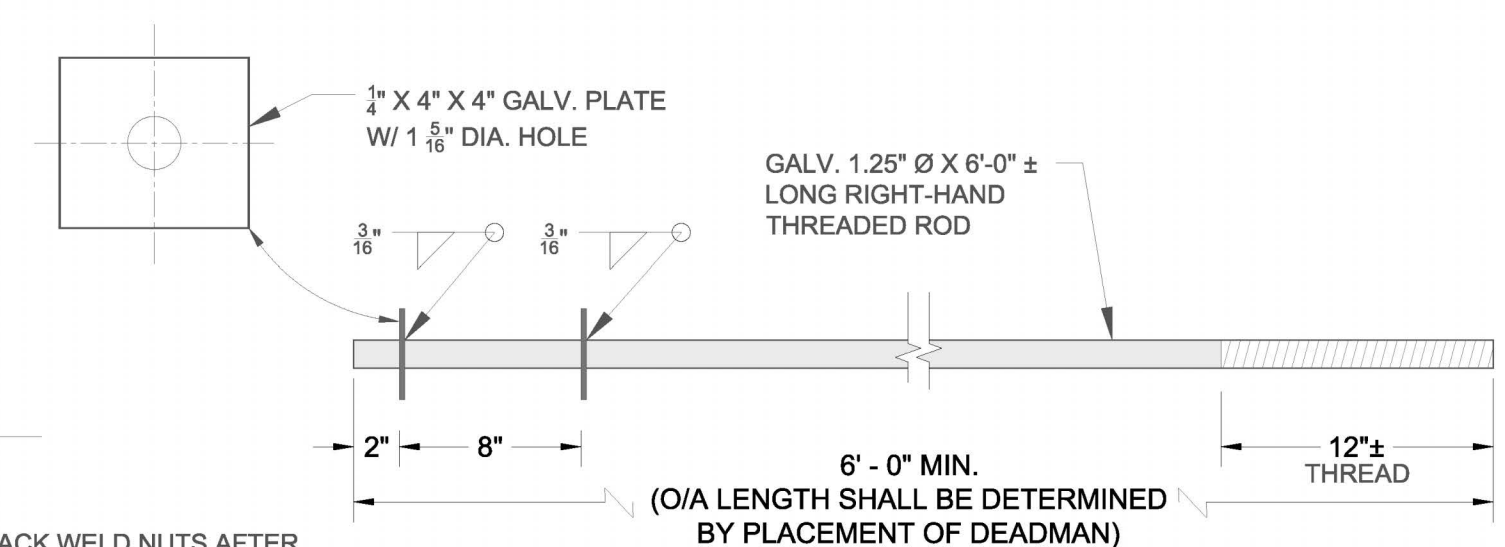
WIND SEIZING WIRE IN A TIGHT SPIRAL AROUND THE ROPE END TO A 1" LENGTH OF SEIZING. TWIST THE TWO ENDS OF THE SEIZING WIRE AT THE CENTER OF THE SEIZING LENGTH. USE ZINC COATED STEEL WIRE.



**4 CABLE ANCHORAGE DETAIL @ PULL & ANCHOR POST - PUBLIC SIDE**

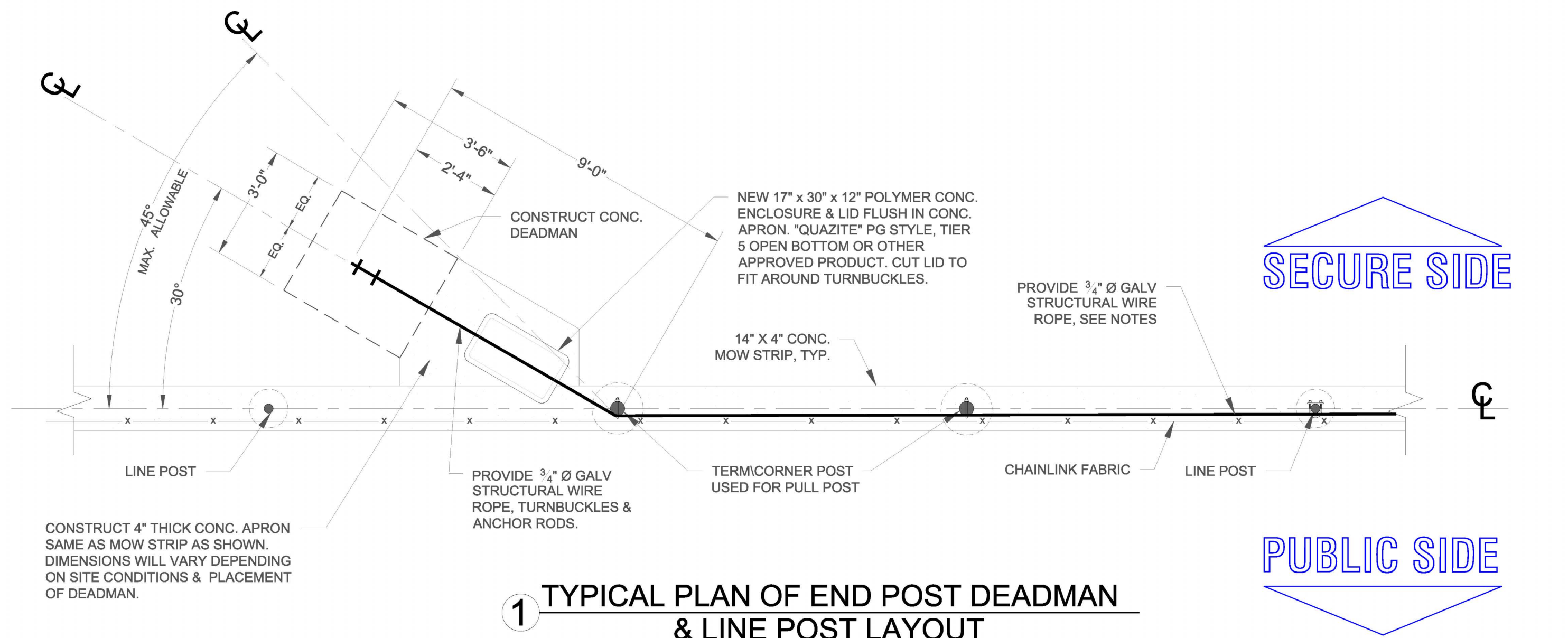


**2 DEADMAN & CABLE TERMINATION**

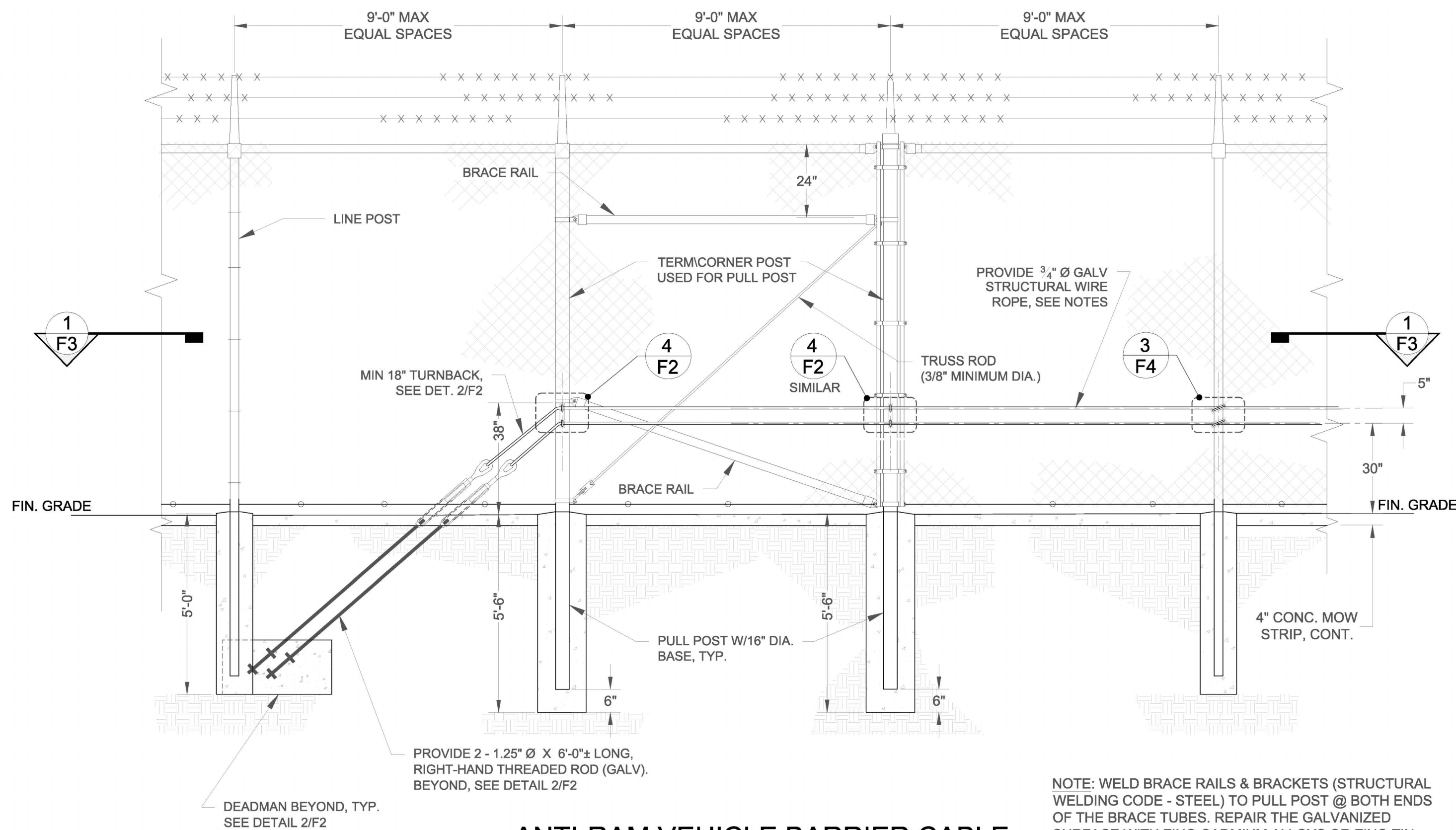


**5 DEADMAN ANCHOR ROD**





1 TYPICAL PLAN OF END POST DEADMAN & LINE POST LAYOUT



2 ANTI-RAM VEHICLE BARRIER CABLE PUBLIC SIDE ELEV

NOTE: WELD BRACE RAILS & BRACKETS (STRUCTURAL WELDING CODE - STEEL) TO PULL POST @ BOTH ENDS OF THE BRACE TUBES. REPAIR THE GALVANIZED SURFACE WITH ZINC-CADMIUM ALLOYS OR ZINC-TIN LEAD ALLOY "REPAIR STICKS."

## NOTES:

1. BASE BID INCLUDES THIS WORK ONLY IN TWO AREAS DESIGNATED ON SHT T2.
2. BID OPTIONS: MAY BE ACCEPTED FOR THE REMAINING RUN OF NEW FENCE AS SHOWN WITH GREEN LINE SYMBOL ON SHT. T2 & NOTE 7.
3. DRAWINGS SHOW ONE TERMINAL END OF THE ANTI-RAM VEHICLE BARRIER CABLE RUN. CONSTRUCT THE OPPOSITE END THE SAME OR SIMILAR WITH LITTLE DEVIATION. EXPECT TO MAKE ADJUSTMENTS FOR SITE CONDITIONS.
4. REINFORCING CABLES SHALL BE U.S. DOMESTIC MINIMUM 3/4" Ø 6x19 CLASS WIRE ROPE, EXTRA IMPROVED PLOW STEEL (EIP), INDEPENDENT WIRE ROPE CORE (IWRC), CONFORMING TO ASTM A1023 AND GALVANIZED IN ACCORDANCE WITH ASTM A475 CLASS A, & HAVE A MINIMUM BREAKING STRENGTH OF 40,000 POUNDS (20 TONS).
5. CABLES SHALL BE CONTINUOUS FROM DEADMAN-TO-DEADMAN AT OPPOSITE ENDS OF THE RUN. NO CABLE SPLICES ALLOWED. CABLE BARRIER SHALL BE INSTALLED BETWEEN FENCE POST AND FENCE FABRIC ON THE PUBLIC SIDE AS SHOWN. U-BOLTS SHALL BE INSTALLED PERPENDICULAR TO THE LAY OF THE STRANDS OF THE WIRE ROPE AND SHALL BE TIGHTENED AFTER SAG IN CABLE BARRIER HAS BEEN REMOVED. UNLESS INDICATED OTHERWISE, CONCRETE DEADMAN SPACING SHALL BE AT MAXIMUM 200' INTERVALS & TURNING POINTS (CORNERS).
6. DEADMEN SHALL BE INSTALLED ON THE SECURED (INTERIOR PROTECTED) SIDE OF THE FENCE. CABLES SHALL BE INSTALLED ON THE EXTERIOR SIDE OF POST UNDER THE FENCE FABRIC.
7. CONCRETE DEADMEN SHALL CURE A MINIMUM OF 7 DAYS UNDER ACCEPTABLE AMBIENT CONDITIONS AS APPROVED BY THE GOVERNMENT PRIOR TO APPLYING LOADS - STRAIGHTENING & TENSIONING CABLES.
8. ALL STRUCTURAL STEEL (INCLUDING HARDWARE) USED IN CABLE ANCHORING SYSTEM SHALL BE HOT-DIPPED GALVANIZED. ANY AREAS WHERE COATING IS DAMAGED OR REMOVED SHALL BE TOUCHED UP WITH ZINC RICH COATING.
9. TURNBUCKLES SHALL BE 1 1/4"x18", TYPE I, GALVANIZED IN ACCORDANCE WITH ASTM F1145.
10. WIRE ROPE CLAMPS SHALL BE TYPE I, GALVANIZED, AND CONFORM TO FS FF-C-450.
11. ALL THREADED RODS, U-BOLTS, AND BOLTS SHALL CONFORM TO ASTM A307 AND SHALL BE INSTALLED WITH F844 WASHERS AND A563 NUTS. ENTIRE BOLT ASSEMBLY SHALL BE GALVANIZED IN ACCORDANCE WITH ASTM A153. INSTALL PER MANUFACTURER'S RECOMMENDATIONS.
12. ZINC RICH COMPOUND FOR REPAIRS SHALL BE 95% METALLIC ZINC, BY WEIGHT IN DRIED FILM; INSTALL AT LEAST TWO COATS, 4 MILS MIN TOTAL THICKNESS.
13. ALL CONCRETE WORK SHALL BE IN ACCORDANCE WITH THE LATEST EDITION OF ACI 318. CONCRETE SHALL HAVE A MINIMUM 28-DAY COMPRESSIVE STRENGTH OF 4000 PSI.
14. THREADED RODS SHALL BE 1 1/4", GALVANIZED. IN ACCORRDANCE WITH ASTM A307 OR A36.
15. WELDED BRACE RAILS SHALL BE INSTALLED AS PER THE PLANS AND AWS D1.1 (STRUCTURAL WELDING CODE - STEEL) WHERE WELDING IS REQUIRED. DAMAGE TO GALVANIZED SURFACES SHALL BE REPAIRED WITH "REPAIR STICKS" OF ZINC-CADMIUM ALLOYS OR ZINC-TIN LEAD ALLOYS AS PER AWS-WZC D19.



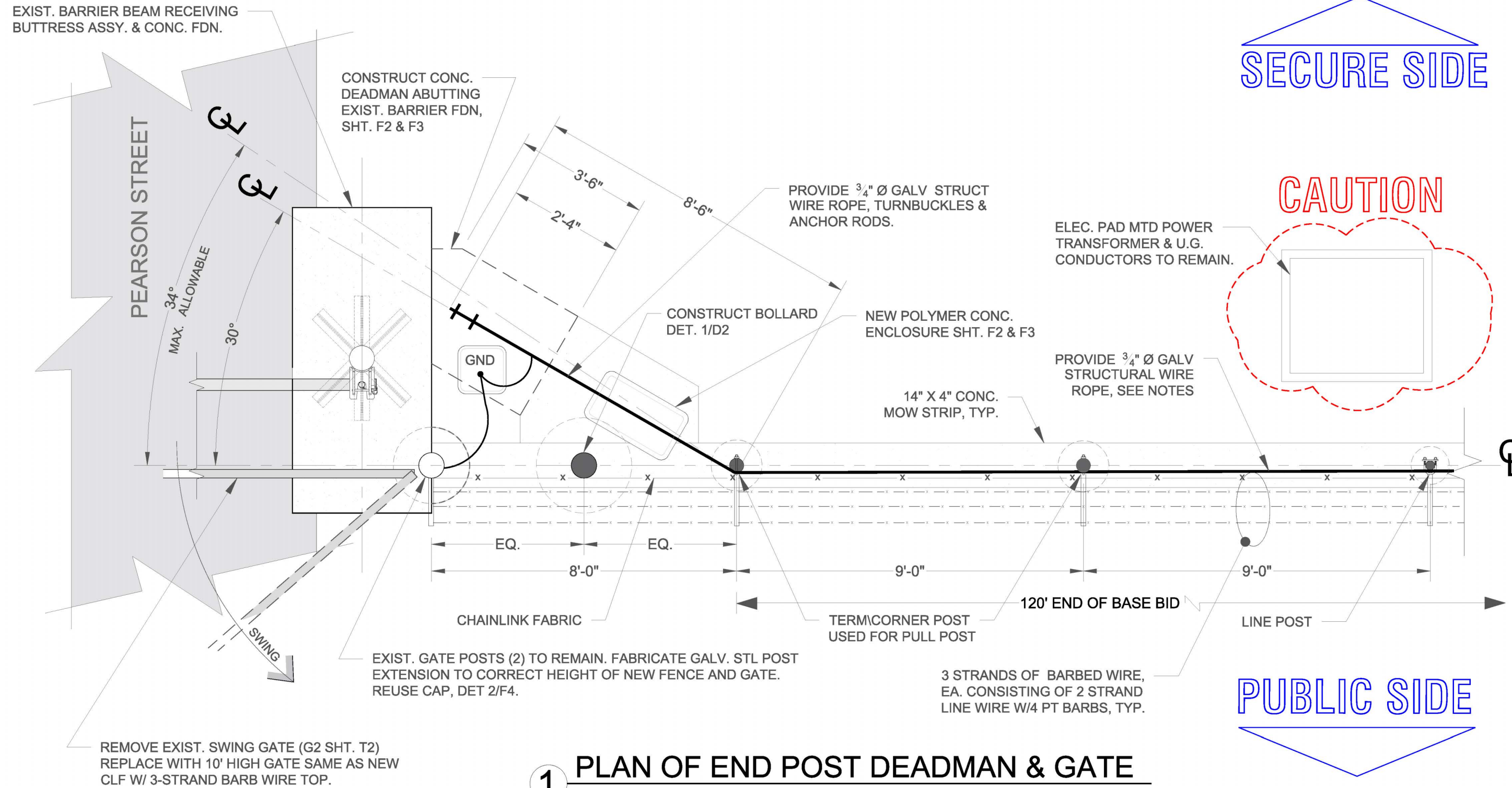
NORTH



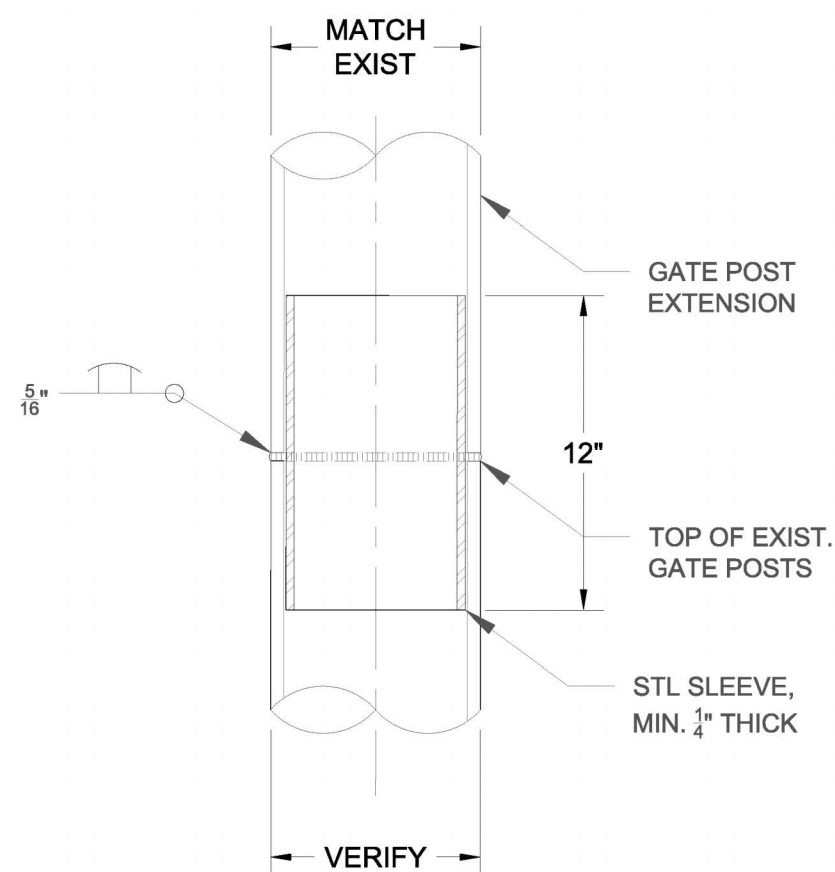
SECURE SIDE

CAUTION

PUBLIC SIDE

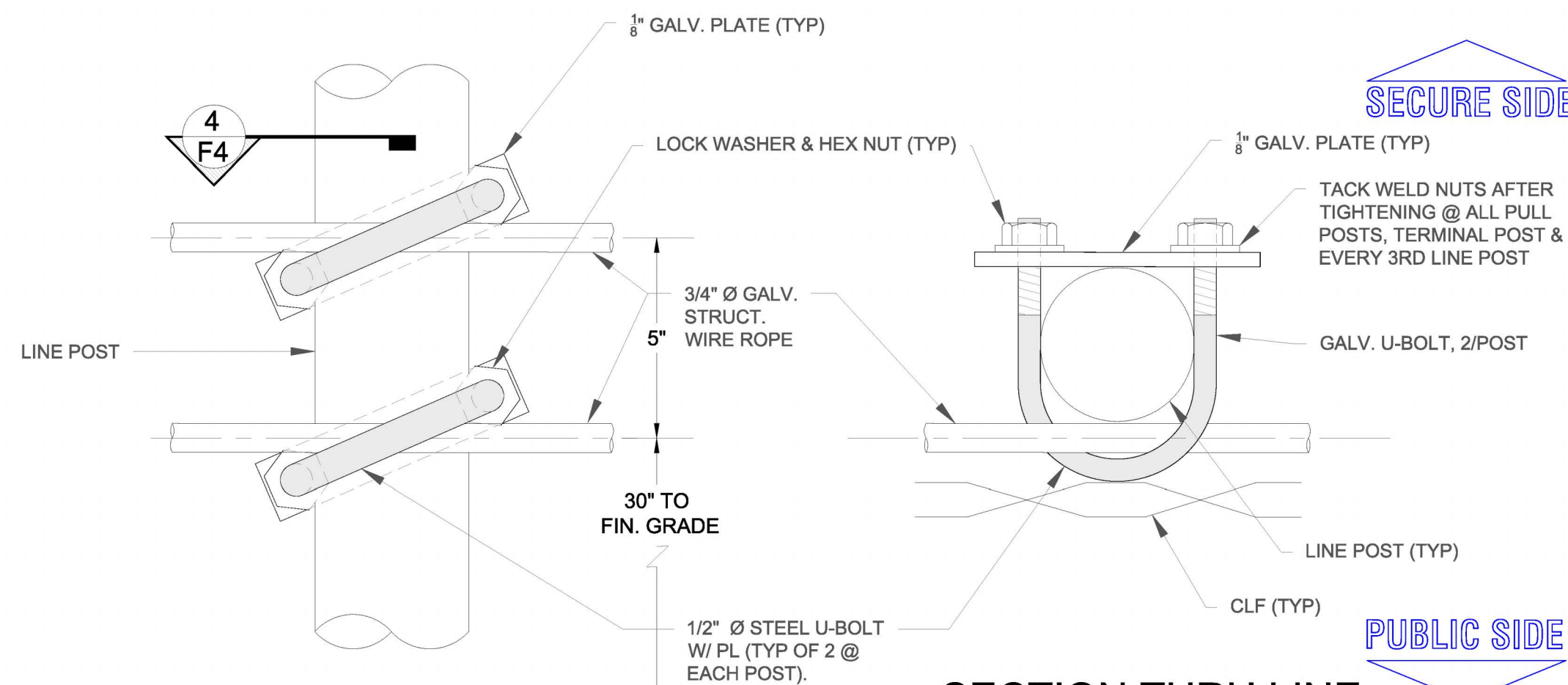


## 1 PLAN OF END POST DEADMAN & GATE @ G2, PEARSON STREET



## 2 SECTION THRU POST EXTENSION TYPICAL

NOTE: REPAIR ALL GALV. SURFACES WITH ZINC-CADMIUM ALLOYS OR ZINC-TIN LEAD ALLOY COATING (SEE NOTE 8, F3).



## 3 LINE POST BARRIER CABLE CONNECTION

## 4 SECTION THRU LINE POST

### NOTES:

1. BASE BID : ALL WORK ON THIS SHEET IS INCLUDED IN THE BASE BID. SEE SHT. T2 FOR OVERALL PLAN AND SHT. F3 NOTES.
2. DRAWINGS SHOW ONE TERMINAL END OF THE ANTI-RAM VEHICLE BARRIER CABLE RUN. CONSTRUCT THE OPPOSITE END THE SAME OR SIMILAR WITH SOME DEVIATION. EXPECT TO MAKE ADJUSTMENTS FOR SITE CONDITIONS.
3. SEE NOTE 10 - SHT. T2 FOR BOLLARDS ACROSS PEARSON STREET ON THE WEST END OF GATE G2 ON THE SECURE SIDE.
4. SEE SHT. C5 FOR ELECTRICAL GROUNDING REQUIRED. FURNISH & INSTALL IN-GROUND ELEC. BOX, GROUND ROD & CONDUCTORS. BOND THE FENCE, CORNER POST & DEADMAN THREADED RODS TO THE GROUND (GND) ROD IN QUARTZITE BOX WHERE SHOWN.

MAY 2020



DRAWN BY: DJW

XGFG 192006  
UPGRADE MAIN GATE BARRIERS

## NEW ANTI-RAM VEHICLE BARRIER CABLE

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WISCONSIN AIR NATIONAL GUARD  
115<sup>TH</sup> FIGHTER WING  
TRUAX FIELD AIR NATIONAL GUARD BASE  
3110 Mitchell Street  
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608 245-4571

F4





## WISCONSIN DEPARTMENT OF NATURAL RESOURCES NOTICE OF FINAL GUIDANCE & CERTIFICATION

Pursuant to ch. 227, Wis. Stats., the Wisconsin Department of Natural Resources has finalized and hereby certifies the following guidance document.

### DOCUMENT ID

EA-19-0001

### DOCUMENT TITLE

Wisconsin PFAS Aqueous (Non-Potable Water) and Non-Aqueous Matrices Method Expectations

### PROGRAM/BUREAU

Certification Services / Environmental Analysis & Sustainability

### STATUTORY AUTHORITY OR LEGAL CITATION

Wis. Stats. s. 299.11 and Wis. Admin. Code s. NR 149.41 (2)

### DATE SENT TO LEGISLATIVE REFERENCE BUREAU (FOR PUBLIC COMMENTS)

9.16.19

### DATE FINALIZED

12.16.19

### DNR CERTIFICATION

*I have reviewed this guidance document or proposed guidance document and I certify that it complies with sections 227.10 and 227.11 of the Wisconsin Statutes. I further certify that the guidance document or proposed guidance document contains no standard, requirement, or threshold that is not explicitly required or explicitly permitted by a statute or a rule that has been lawfully promulgated. I further certify that the guidance document or proposed guidance document contains no standard, requirement, or threshold that is more restrictive than a standard, requirement, or threshold contained in the Wisconsin Statutes.*

Signature

Date

12/10/2019





## Wisconsin PFAS Aqueous (Non-Potable Water) and Non-Aqueous Matrices Method Expectations



- Version 12.16.2019 -

Per- and Polyfluorinated Alkyl Substances (PFAS) Analysis Using Isotope Dilution by LC/MS/MS

The purpose of this document is to provide the expectations that will help the Program determine if a laboratory's method is considered suitable for analysis of PFAS in aqueous (non-potable water) and non-aqueous matrices for Wisconsin.

The Program has the legal authority under NR 149.41 (2) to determine whether the method selected by a laboratory is suitable for the matrix, type of analyte, expected level of analyte, regulatory limit, and anticipated interferences in the sample, when methods are not prescribed by covered programs under NR 149 or permits issued by the department.

Once the EPA publishes their 1600 series isotope dilution method, the Program will defer to that method for certification.

Potable water samples are analyzed utilizing EPA 537.1.

{F} = when "{F}" is listed after an expectation and the expectation is not met, then qualify the associated results on the test report. The qualifier can refer the data user to the narrative where detail is provided that indicates what the non-conformance was, and if known, the possible effects on the sample results.

Definitions are provided in Section X, "Definitions," of this document.

### I. Sample Handling

1. Instruct sample collectors to collect grab samples in high density polyethylene or polypropylene containers. {F} Avoid polytetrafluoroethylene (PTFE) containers and contact with PTFE surfaces.
2. Instruct sample collectors to collect an equipment blank when using equipment in the field to collect samples. {F}
3. Instruct sample collectors not to fill aqueous sample containers completely.
4. There is no chemical preservation necessary, just temperature preservation. Instruct sample collectors to ship aqueous and solid samples at above their freezing point to 6 °C. {F} Instruct sample collectors to ship tissue samples frozen. {F} Measure and document the temperature of aqueous and solid samples at sample receipt. Tissue samples received frozen can be documented as "frozen" at sample receipt.
5. Store aqueous and solid samples at above their freezing point to 6 °C at the laboratory. {F} Store tissue samples at less than or equal to -10 °C at the laboratory. {F} Store all extracts at 0 – 6 °C at the laboratory. {F}
6. Aqueous and solid sample holding times are within 28 days from collection to extraction and within 30 days from extraction to analysis. {F} Tissue sample holding times are within 1 year from collection to extraction and within 30 days from extraction to analysis. {F}
7. Rinse aqueous sample containers and all extract containers after transfers with one or more rinses of polar solvent to remove any PFAS that may have been adsorbed to container walls.
8. Thoroughly vortex or mix extracts and standards before transfer or aliquoting to remove any PFAS that may have been adsorbed to container walls.
9. Thoroughly vortex autosampler vials before loading the autosampler to remove any PFAS that may have adsorbed to container walls.





## II. Initial Demonstration of Capability (IDC)

1. All analysts performing testing are expected to pass an IDC. If analysts perform only the extraction steps, then they are expected to pass the extraction portion of an IDC. If analysts perform only the analysis steps, then they are expected to pass the analysis portion of an IDC.
2. Analyze standards of all target (native) analytes and extracted internal standards (EIS) to determine retention times of the linear and branched isomers.
3. Analyze a method blank. The results are expected to be less than one-half the method reporting limit (MRL).
4. Assess precision and recovery by performing the entire procedure on four laboratory control samples (LCS) spiked at a midrange concentration of the initial calibration for each target (native) analyte. The average recovery is expected to be within 65-135%, and the RSD is expected to be less than or equal to 30%.
5. Assess recovery of the extracted internal standards (EIS) in each LCS. Except for FOSA, NMeFOSA, NEtFOSA, NMeFOSE, and NEtFOSE, EIS recoveries are expected to be within 50–150%. For FOSA, NMeFOSA, NEtFOSA, NMeFOSE, and NEtFOSE, EIS recoveries are expected to be within 20 – 150%.

## III. Field Quality Control Samples

1. **Equipment blanks** (one per sampling event when equipment is used in the field to collect samples) – The results are expected to be less than the highest of the following {F}:
  - a. 1/2 the MRL
  - b. 1/10 the sample concentration

It is not necessary to qualify equipment blank detections between the MDL and one-half the MRL.

2. **Field blanks** (one per sampling event for each sampling site) – The results are expected to be less than the highest of the following {F}:
  - a. 1/2 the MRL
  - b. 1/10 the sample concentration

It is not necessary to qualify field blank detections between the MDL and one-half the MRL.

3. **Field duplicates** (one per sampling event for each sampling site) – The RPDs are expected to be less than or equal to 30% when analyte concentrations are greater than twice the MRL. {F} The RPDs are expected to be less than or equal to 50% when analyte concentrations are the MRL and twice the MRL. {F}





#### IV. Batch Quality Control Samples

1. **Method blank** (one per batch) – The results are expected to be less than the highest of the following {F}:
  - a. 1/2 the MRL
  - b. 1/10 the sample concentration

It is not necessary to qualify method blank detections between the MDL and one-half the MRL.

Method blanks are processed along with and under the same conditions, including all sample preparation steps (i.e. filtering, centrifuging), as the associated samples in the preparation batch.

2. **Laboratory control sample** (one per batch) – Spike with all target (native) analytes.

Laboratory control samples are processed along with and under the same conditions, including all sample preparation steps (i.e. filtering, centrifuging), as the associated samples in the preparation batch.

For aqueous and solids batches, spike the LCS at a low range (1 – 2x MRL) in each batch, or the laboratory may rotate spike concentrations between three consecutive batches alternating low range, midrange, and high range. Midrange and high range are relative to the initial calibration range. For aqueous and solid batches, the recoveries are expected to be within 60-135%, except for the low range (1 – 2x MRL) where the recoveries are expected to be within 50-150%. {F}

For tissue batches, spike the LCS at midrange. For tissue batches the recoveries are expected to be within 60-135% with the following exceptions: for PFHxDA, PFODA, and NMeFOSA, the recoveries are expected to be within 50-135%; for PFDS, PFDoS, and 4:2 FTS, the recoveries are expected to be within 40-135%. {F}

3. **Extracted internal standards (EIS)** – Spike field samples and all quality control samples (preparation and instrument) with internal standards. The recoveries of these internal standards are used to adjust target (native) analyte concentrations. These isotopically labeled internal standards are added to the sample at the very beginning of the procedure, before extraction, centrifuging, filtering or phase separation takes place.

In order to report quantitative results for the target (native) analytes using the EIS, a minimum signal to noise ratio of 10:1 is expected for each EIS. Do not report results with a qualifier if this minimum is not achieved.

Except for FOSA, NMeFOSA, NEtFOSA, NMeFOSE, and NEtFOSE, the EIS recoveries are expected to be within 25-150% in samples. For FOSA, NMeFOSA, NEtFOSA, NMeFOSE, and NEtFOSE, these EIS recoveries are expected to be within 10-150% in samples. Once enough data points have been collected, the laboratory may develop their own statistical limits for these five EIS in samples. The statistical limits can be different than 10–150% as long as the expected minimum 10:1 signal to noise ratio is maintained for each EIS.

If any EIS recoveries are outside of limits in a sample, reinject the sample. If the EIS recovery fails again, the data may be reported with a qualifier. {F}

Use exact isotopically labeled analogs for the EIS where commercially available. As of December 2019, at least 25 of the 36 PFAS for which Wisconsin is offering certification are available as exact isotopically labeled analogs of the target (native) analytes. As of December 2019, the following 11 PFAS do not have exact isotopically labeled analogs commercially available and are therefore not currently necessary: PFTriA, PFODA, PFPeS, PFHpS, PFNS, PFDS, PFDoS, 10:2 FTSa, DONA, 9Cl-PF3ONS, and 11Cl-PF3OUDS.





For these 11 PFAS without an exact isotopically labeled analog commercially available, use an alternate EIS. The alternate EIS is expected to be isotopically labeled and is expected to be a chemically similar analyte that is close in retention time to the target (native) analyte. The alternate EIS may be from the same functional group as the target (native) analyte or have the same chain length as the target (native) analyte (whichever gives better performance). Typically, the alternate EIS comes from those EIS that are already in use. The same EIS can be used for more than one target (native) analyte.

### V. Calibration (Initial and Continuing)

1. Perform initial calibration at setup and after an ICV or CCV standard failure. If an ICV or CCV standard fails, the laboratory may immediately analyze two additional consecutive ICV or CCV standards. If either of the two fails, or if immediate analysis is not possible, it is expected that a new initial calibration is performed. If both pass, then sample analysis can continue without a new initial calibration. If a CCV fails high and there are no detections in the associated samples, then analysis can proceed.
2. Initial calibration functions are expected to be as follows:
  - a. Calibration factors have an RSD that is less than or equal to 20%.
  - b. Linear regressions have a coefficient of determination that is greater than or equal to 0.99 and use a minimum of five non-zero concentration standards.
  - c. Quadratic regressions have a coefficient of determination that is greater than or equal to 0.99 and use a minimum of six non-zero concentration standards.
  - d. Do not force linear and quadratic regressions through zero.
  - e. For each calibration standard, reprocess the target (native) analyte against the chosen calibration function. The reprocessed recoveries are expected to be within 70–130% of their actual concentrations, except for the lowest concentration standard, whose reprocessed recoveries are expected to be within 50–150% of their actual concentrations.
3. It is expected that sample analysis is not performed if the initial calibration fails.
4. Analyze standards of all target (native) analytes and EIS to determine retention times of the linear and branched isomers. Analyze branched isomers that have commercially available standards. As of December 2019, the following PFAS are commercially available as branched isomer analytical (quantitative) standards: PFHxS, PFOS, NMeFOSAA, and NEtFOSAA. As of December 2019, PFOA is commercially available as a branched isomer technical grade (qualitative) standard.
5. When an initial calibration is performed, it is expected that the midrange standard is used to establish absolute retention times. When an initial calibration is not performed, it is expected that the first CCV is used to establish absolute retention times.
6. Retention times of the target (native) analytes and the EIS are expected to fall within 0.4 minutes of the established absolute retention times. Comparison of the target (native) analyte and EIS retention times can help determine if analyte shifts occurred due to matrix effects.
7. **ICV (2<sup>nd</sup> source)** – It is expected that the ICV is performed with each new initial calibration before sample analysis. The ICV is analyzed after the ICB. As of December 2019, the following PFAS may be difficult to find as second sources and are therefore not currently necessary: PFHxDA, PFODA, PFDoS, NMeFOSA, NEtFOSA, NMeFOSE, and NEtFOSE. Recoveries in the ICV are expected to be within 70-130%. It is expected that sample analysis is not performed if the ICV fails.





8. **ICB** – It is expected that the ICB is analyzed immediately after the highest standard in the initial calibration and before the ICV to demonstrate the instrument is free from levels of contaminants that would bias results. The results of the ICB are expected to be less than one-half the MRL.
9. **CCV** – It is expected that CCVs are performed at the beginning and end of each analysis batch and after every 10 field samples.
  - a. It is expected that the concentrations in the first CCV on non-initial calibration days are at the MRL.
  - b. Target (native) analyte recoveries are expected to be within 50-150% for the CCV analyzed at the MRL.
  - c. Target (native) analyte recoveries for all other CCVs are expected to be within 70-130%.
  - d. It is expected that samples results are only reported when bracketed by passing CCVs unless the recovery failure is high and there are no detections of that analyte in the associated samples.
10. **CCB** – It is expected that the CCB is analyzed immediately after each CCV to demonstrate the instrument is free from levels of contaminants that would bias results. If method blanks or reagent blanks are analyzed after a CCV instead of a CCB, then it is expected that the CCB limits are used for assessment. The results of the CCBs are expected to be less than one-half the MRL.
11. It is expected that the same EIS as those used in samples are added to the initial calibration standards, ICV, CCVs, ICBs, and CCBs at the same concentration used in samples. The calibration standards (initial and continuing) are not extracted like samples. Since there is no matrix effect or extraction performed on these instrument quality control samples, the recoveries of the EIS are expected to be within 50 – 150%.

### VI. Aqueous Sample Extraction

1. Extract the entire sample received in the sample container in which it was collected unless the exceptions listed below apply.
  - a. Samples received at extremely high PFAS concentrations may be subsampled. {F}
  - b. If more sample volume is received than what can be extracted through the solid phase extraction (SPE) cartridge, then subsampling is allowed. {F}

Adsorption of target (native) analytes to sample collection container walls is known to occur in aqueous samples. Extract the entire aqueous sample volume. Subsampling of aqueous samples from the sample collection container is discouraged and can result in significant loss of longer-chain PFAS (e.g. carboxylic acids  $\geq$  C9, sulfonic acids  $\geq$  C7).

2. Spike the sample in the sample bottle it was received in by adding the EIS. Cap, invert and mix. It is expected that the EIS that are spiked into the sample are provided sufficient time to equilibrate in the sample before further processing. This allows the EIS time to disperse proportionally into the liquid phase and solid phase – same as the target (native) analytes and thereby providing a more accurate result. Add the EIS before any extraction, centrifuging, filtering or phase separation takes place.

Biphasic and problematic sample matrices may have to use a different spiking procedure. It is best for the laboratory to contact the client prior to spiking and extraction to determine the best course of action to meet their data quality objectives. In these events, include detail in the narrative as to why spiking into the sample bottle was not possible, what was done instead, and if known, the possible effects on the sample results. {F}





3. If particulates in the sample have to be removed before using SPE, centrifuge the sample and take the liquid phase through the SPE. Samples should only be centrifuged when the suspended solids content visually appears to be high enough, by chemist inspection, that it would cause the SPE cartridge to clog.

The laboratory could consider creating a “percent solids reference sample” that would include the minimum solids the laboratory has tested that would clog the SPE cartridge and use it to compare it to field samples. For reference, the Department of Defense has indicated that samples with percent solids greater than one percent may require centrifuging before performing the SPE procedure. Ideally, the entire sample is extracted, including the suspended solids.

4. If aqueous samples with a solid phase are centrifuged, the solid phase of the sample is expected to be a plug at the bottom of the container. It is expected that the solid phase remains in the container when rinsing the container walls with the polar elution solvent. Rinsing the container walls would therefore also include rinsing of the solids. If the polar elution solvent disrupts the solid phase significantly, the container can be centrifuged again before removing the solvent for use during the elution step of the SPE procedure.
5. If a total sample concentration is needed and there are significant solids in the sample, the initial spike of EIS into the sample container is sufficient for both phases. There is no need to re-spike the solid phase with EIS if it is being extracted separately.
6. Using filters to separate the solid phase from the liquid phase is discouraged unless there is data to demonstrate that the filters used do not result in contamination greater than one-half the MRL.
7. In the cases where a filter is used to separate the solid phase from the liquid phase, it is expected that the filter would also be rinsed to remove any potentially adsorbed PFAS. The filtrate is then added to the SPE cartridge during the elution step.
8. The data quality objectives from the data user should determine whether the solid phase of the sample has to be extracted or not. Not analyzing the solid phase may lead to a low bias in total sample concentration. Analyzing the liquid phase only would provide a liquid sample concentration result. It is expected that the laboratory would make it clear to the data user whether the reported concentrations are a total or liquid concentration sample result.
9. Determine sample volume by marking the sample level on the bottle or by weighing. It is expected that sample volumes would not be measured with a graduated cylinder. Sample volumes are expected to be measured and not assumed by container size.

When the sample has significant solids, the laboratory should account for the weight or volume displaced by the solids in the initial sample volume determination and include this information in the test report.

10. Use an appropriate SPE cartridge for the target (native) analytes reported. A weak anion exchange cartridge has been shown to work with the PFAS for which Wisconsin is offering certification.
11. One or more rinses of polar solvent can be used for quantitative transfers. Rinse the sample bottle and cap with elution solvent, pour the solvent from each rinse through the SPE cartridge during the elution step, and collect the filtrate for analysis.
12. Bring to a quantitative final volume with the final injection solvent and vortex well.





## VII. Non-Aqueous Sample Extraction

1. Homogenize the entire solid sample received in the sample container in which it was collected in by stirring the solids with a clean spatula or other suitable implement. This would help ensure that a representative subsample is taken.
2. For tissues (e.g. fish, wildlife), the target tissue (liver, fillet, whole fish) is isolated from the rest of the tissue sample. The target (isolated) tissue is ground and is typically provided to the analyst as a subsample. At the time of sample preparation, the analyst is to further homogenize the subsample by stirring with a clean spatula or other suitable implement. This would help ensure that a representative subsample is taken.
3. Spike a portion of the homogenized subsample by adding the EIS directly onto the sample. It is expected that the solvent used to carry the EIS spike onto the sample be allowed to evaporate prior to addition of the extraction solution.
4. Extract the PFAS from the non-aqueous samples with an appropriate solution prior to clean-up.
5. Use an appropriate clean-up cartridge (i.e. ENVI-Carb, W-AX, ...) to remove the organic analytes extracted from the soil matrix. More than one type of clean-up cartridge can be used.
6. Use a clean-up cartridge on the fish tissue extract to eliminate known interferences with PFOS (e.g. bile acids such as taurodeoxycholic acid (TDCA)).
7. Ensure that all transfers are quantitative by solvent-rinsing with the elution solvent.
8. Bring to a quantitative final volume with the final injection solvent and vortex thoroughly.

## VIII. Sample Analysis

1. Use an LC/MS/MS that is capable of negative ion ESI, produces unique product ions within retention time windows, and is able to provide a minimum of 10 scans across each peak.
2. Perform mass calibration such that the range of masses associated with all precursor and product ions are bracketed for both the primary and confirmation transitions. Documentation is expected to be available to demonstrate that the mass calibration covers this range. Calibrate the mass scale using the calibration analytes and procedure from the instrument manufacturer.
3. Analyte identification is performed using retention times, Signal/Noise ratio, Quantitation Parent Ion to Quantitation Daughter Ion (Quantitation Ion Transition), Confirmation Parent Ion to Confirmation Daughter Ion (Confirmation Ion Transition) and the Ion Transition Ratio.
4. Calculate sample results for the target (native) analytes that have exact isotopically labeled standards using isotope dilution (recovery correction using the EIS).
5. Calculate sample results for the target (native) analytes that do not have exact isotopically labeled standards using an alternate extracted isotopically labeled standard and internal standard quantitation recovery correction (recovery correction using the alternate EIS).
6. Use analytical (quantitative) standards containing both branched and linear isomers where commercially available. The analytical branched isomer standards are included in the initial calibration the same as the linear isomer





standards. Branched isomers in samples are quantitated against these analytical branched isomer standards. To calculate the target (native) analyte result, sum the resulting concentrations of all branched and linear isomers that have corresponding analytical standards.

- Where analytical standards are not available for the branched isomers, use qualitative (technical grade) standards to identify the branched isomer using retention times, transitions, and ion transition ratios. Quantitate target (native) analytes that use qualitative branched isomer standards by integrating the branched and linear isomer peaks and sum the peak areas to get a total area. Calculate the target (native) analyte concentration using the linear isomer.

Do not include branched isomer peaks in the initial calibration when qualitative standards are used, and do not use calibration functions from the qualitative branched isomer standards to quantitate branch isomer concentrations.

- It is expected that the target (native) analytes that have exact labeled analogs would elute within 0.1 min of their analogs. {F}
- Have a written policy on how retention time windows are established.
- It is expected that the method reporting limit (MRL) concentration would not be below the lowest standard concentration in the initial calibration.
- The MDL is expected to be less than the MRL.
- Report sample results and all quality control blank results to the MDL and include the MRL with each result. Qualify results reported between the MDL and MRL as estimated concentrations.

Example 1: MDL = 0.6, MRL = 2, sample result = 0.4. Report as:

<u>Result</u>	<u>MDL</u>	<u>MRL</u>
<0.6	0.6	2.0

Example 2: MDL = 0.6, MRL = 2, sample result = 0.8. Report as:

<u>Result</u>	<u>MDL</u>	<u>MRL</u>
0.8 J	0.6	2.0

- The MDL for PFOS and PFOA in non-potable waters are each expected to be no higher than 2 ng/L.
- It is expected that high density polyethylene or polypropylene autosampler vials are single injection use only unless they are immediately recapped.
- It is expected that all sample results are reported from a response that is no higher than the highest response in the initial calibration, except for samples that saturate the instrument. If supplemental EIS is needed to quantitate dilutions, qualify the results that used the supplemental EIS (in this case, true isotope dilution was not achieved).
- It is expected that sample results that saturate the instrument are reported with "E" flags. {F}
- For target (native) analytes, the Signal to Noise (S/N) ratio is expected to be greater than or equal to 3:1 for quantitation ions and confirmation ions. If the S/N is not achieved, it is expected that the peak would not be used in any way and the analyte would be reported as "not detected."





18. All analytes that have two transitions are expected to include two transitions ions in the analysis (precursor ion to quantitation ion and precursor ion to confirmation ion). Use the confirmation ion for positive analyte identification. The department has provided a list of target (native) analytes and confirmation ions in section **XII, “Wisconsin Laboratory Accreditation Program PFAS Certification Offerings with Ions,”** of this document.

19. Assess primary and secondary ion transition ratios. It is expected that recoveries be within 50–150% of the value calculated from the midrange standard in the ICAL on ICAL days or from the beginning CCV on non-ICAL days. {F}

The transition ratio =  $\frac{\text{quantitation ion abundance}}{\text{confirmation ion abundance}}$  or  $\frac{\text{confirmation ion abundance}}{\text{quantitation ion abundance}}$

Either ratio protocol presented above can be used, but it is expected that the protocol is consistently used for all analytes.

When the ion ratio fails, it is expected that the target (native) analytes would still be reported but qualify them as failing the ion ratio. {F} The ion transition ratio can help identify if bias is present. Ratios can be outside of limits due to interferences or the presence of branched isomers that are in the sample but not in the quantitation standards.

20. Document the primary and confirmation transitions and the ion transition ratio.

21. It is expected that the following transitions are used for quantitation of the following analytes [precursor – product] unless a technically justified reason is used and documented:

- a. PFOA 413-369
- b. PFOS 499-80
- c. PFHxS 399-80
- d. PFBS 299-80
- e. 4:2 FTS 327-307
- f. 6:2 FTS 427-407
- g. 8:2 FTS 527-507
- h. NEtFOSAA 584-419
- i. NMeFOSAA 570-419

22. The laboratory is expected to determine at what concentration the instrument has carryover at concentrations greater than one-half the MRL. The laboratory is expected to have a documented procedure to bring the instrument back in control after encountering a sample with carryover. PFAS have demonstrated a delayed release in the system.

23. Report results in acid form.

24. Verify standard purity and ensure that any standards with less than 98% purity are corrected for in the calculations.

25. Mass correct salt content in all calibration standards purchased as salts.

26. Perform a moisture analysis on solid samples (on a subsample different than that used for extraction) and adjust the final concentration of solid samples for the percent moisture.

27. If only the liquid phase of a biphasic sample was extracted, report the results as liquid concentration results instead of total sample concentration results. The lab should report the weight of the solid phase not prepared in this case. This can be detailed in the narrative.





28. If the data quality objective is to obtain a total sample concentration and the sample is biphasic, then extract and analyze both phases.
29. Do not subtract quality control blank values from sample result values.
30. Integrate linear and branched isomers in the samples in the same manner as the standards.
31. Include the following elements in the laboratory SOP:
  - a. The extracted internal standards used to calculate the result of each target (native) analyte reported.
  - b. The mass used for the precursor ion for each analyte.
  - c. The mass used for the product quantitation ion for each analyte.
  - d. The mass used for the product confirmation ion for each analyte.
  - e. Instructions for conditioning and elution of the SPE cartridge.
  - f. Indicate which branched isomers are calculated using the linear isomer standard.
32. PFOA and PFOS WP PT samples are necessary for aqueous (non-potable water) certification of PFOA and PFOS. To obtain the 36-analyte group for aqueous (non-potable water) or non-aqueous from Wisconsin, analyze a PT with a minimum of 6 PFAS that include PFOA and PFOS. It is expected that 80% of the spiked analytes pass.
33. Requirements in NR 149 still apply to this analysis unless otherwise specified in this document.

***AS NEW INFORMATION IS PROVIDED BY THE EPA, THIS DOCUMENT WILL BE UPDATED.***





## IX. Other Considerations

1. Screen a separate aliquot of sample received prior to preparation of a quantitative analysis.
2. Prior to any quantitative analysis, at least one, if not multiple instrument blanks should be analyzed to assess the system for potential contamination. These instrument blanks should include EIS to enable quantitation of the contamination.
3. Evaluate all containers, water, reagents, solvents, materials, SPE cartridges, and equipment as sources of contamination. The lab should be able to demonstrate that these items are not introducing unacceptable positive or negative bias.
4. Supplies should be tested on a lot-by-lot basis.
5. Avoid contact with glassware.
6. Avoid any Teflon including Teflon lined caps.
7. Flush water purification system with 3 liters of reagent water before using.
8. Use LC PEEK tubing and stainless-steel frits.
9. Use polypropylene transfer lines.
10. Replace mobile phase after 48 hours of preparation.
11. Store standards in the containers they were received in and at the storage conditions recommended by the manufacturer.
12. Store solid PFSA standards in a desiccator as they can hydrate over time.
13. PFCA standards in methanol solution may undergo esterification to methyl esters. Ideally, purchase PFCA standard solutions in methanol that contain four mole equivalents of NaOH. Use basic methanol (0.3%  $\text{NH}_4\text{OH}$  v/v in methanol) rather than straight methanol for all standard dilutions to avoid this potential problem.
14. PFSA standards that are  $^{18}\text{O}$ -labelled may exchange with water and therefore reducing purity.
15. To establish retention times, analyze individual standards of each analyte. Analyze a mixed standard of all analytes to confirm their separation and identification.
16. Validate each individual standard and labeled standard by analysis to confirm its identity and the absence of significant impurities.
17. Certified standards have been known to vary by as much as 20% between vendors. The laboratory should be able to demonstrate that the standards being used are of known and defensible quality.
18. Some certified standards are less than 90% pure and often contain impurities that are other PFAS being analyzed.
19. EIS should be 96% or greater purity. When the impurity consists of an unlabeled analyte, the EIS can result in a background artifact that is present in every sample, standard, and blank if the EIS is spiked at excessive concentrations.
20. Different certified standards can have different isomer content.
21. Calibration standards are solvent based only. Matrix matched calibration standards (such as those that include sand or fish tissue) should not be used for isotope dilution methods.
22. If the site where samples are being collected is considered a “newer” spill and source apportionment is one of the data quality objectives, ship the samples with dry ice. PFAS transformation can occur if the samples are not frozen.
23. Although matrix spikes and matrix spike duplicates (MS/MSDs) are not necessary, analyzing them would help with assessing measurement bias for those target (native) analytes that do not have exact labeled isotope analogs.
24. Solid samples should not be air dried unless required by a QAPP.
25. Perform solid and fish tissue PT samples.





## X. Definitions

**Confirmation Ion** - one of the fragment ions (product ions) used to help qualitatively confirm presence of the analyte. The product ion chosen is typically one of the remaining ions with high sensitivity and minimum interferences, after the quantitation ion has been chosen. Not all precursor ions provide confirmation ions.

**Extraction batch** – a set of one to 20 environmental samples of the same certification matrix with a maximum time of 24 hours between the start of processing of the first and last samples in the batch.

**Extracted Internal Standards (EIS)** - isotopically labeled internal standards that undergo the same extraction and analysis as the other analytes in the sample. The EIS are added to the sample at the very beginning of the procedure before extraction, centrifugation, filtering, or phase separation. Ideally, these are exact isotopically labeled analogs of the target (native) analyte so that identical behavior can be assumed. The recoveries of these standards are used to adjust the target (native) analyte results.

**Internal Standard Dilution Quantitation** - measurement of native analytes using an alternate analog (surrogate) isotope (one that has the same chemical behavior and is close in retention time to the native analyte) thus providing a close approximation of matrix effects and losses that can occur during the preparatory and analytical procedures. The native analyte concentration is adjusted for the recovery of the alternate analog isotope. An alternate analog isotope is typically used when an exact analog isotope is not available.

**Method Detection Limit (MDL)** – the minimum measured concentration of a substance that is reported with 99% confidence that the measured concentration is distinguishable from method blank results. The MDL is generated according to the procedure specified in the latest revision of 40 CFR Part 136, Appendix B. The MDL is expected to meet S/N ratio, ion transition ratio, and both quantitation and confirmation ions.

**Method Reporting Limit (MRL)** – the minimum concentration reported as a quantitative value for a method analyte in a sample following analysis. This defined concentration is expected to be no lower than the concentration of the lowest calibration standard for that analyte and is only used if the recovery in the lowest standard is within 50 – 150%.

**Native Analyte** - the analyte being tested in the matrix of interest. It is also the analyte for which a result would be reported. It is defined as native to distinguish it from analyte standards added during the test procedure. Native analyte is also referred to as “target analyte” or “reported analyte.”

**Precursor Ion** – the deprotonated molecule of the analyte. The precursor ion is mass selected and fragmented to produce distinctive product ions of smaller  $m/z$ .

**Product Ion** – one of the fragment ions produced from the precursor ion.

**Quantitation Ion** – one of the fragment ions (product ions) used to quantitate analyte concentrations. The product ion chosen is typically one of high sensitivity and minimum interferences.

**True Isotope Dilution Quantitation** – measurement of native analytes using an exact analog (surrogate) isotope of the native analyte thus eliminating differences in chemical behavior. The native analyte concentration is adjusted for the recovery of the exact analog isotope that has been included in the preparatory and analytical procedures.





## XI. Wisconsin Laboratory Accreditation Program PFAS Certification Offerings – 5.1.19

#	Acronym	Name	CAS #	# carbons	Acronyms (other)
<b>Carboxylic Acids</b>					
1	PFBA	Perfluorobutanoic acid	375-22-4	4	
2	PFPeA	Perfluoropentanoic acid	2706-90-3	5	
3	PFHxA	Perfluorohexanoic acid	307-24-4	6	
4	PFHpA	Perfluoroheptanoic acid	375-85-9	7	
5	PFOA	Perfluorooctanoic acid	335-67-1	8	
6	PFNA	Perfluorononanoic acid	375-95-1	9	
7	PFDA	Perfluorodecanoic acid	335-76-2	10	
8	PFUnA	Perfluoroundecanoic acid	2058-94-8	11	PFUdA, PFUnDA
9	PFDaA	Perfluorododecanoic acid	307-55-1	12	PFDaDA
10	PFTriA	Perfluorotridecanoic acid	72629-94-8	13	PFTriA, PFTriDA
11	PFTeA	Perfluorotetradecanoic acid	376-06-7	14	PFTeDA
12	PFHxDA	Perfluorohexadecanoic acid	67905-19-5	16	
13	PFODA	Perfluorooctadecanoic acid	16517-11-6	18	
<b>Sulfonic Acids</b>					
14	PFBS	Perfluorobutanesulfonic acid	375-73-5	4	
15	PFPeS	Perfluoropentanesulfonic acid	2706-91-4	5	
16	PFHxS	Perfluorohexanesulfonic acid	355-46-4	6	
17	PFHpS	Perfluoroheptanesulfonic acid	375-92-8	7	
18	PFOS	Perfluorooctanesulfonic acid	1763-23-1	8	
19	PFNS	Perfluorononanesulfonic acid	68259-12-1	9	
20	PFDS	Perfluorodecanesulfonic acid	335-77-3	10	
21	PFDoS	Perfluorododecanesulfonic acid	79780-39-5	12	PFDoS
22	4:2 FTSA	4:2 Fluorotelomer sulfonic acid	757124-72-4	6	
23	6:2 FTSA	6:2 Fluorotelomer sulfonic acid	27619-97-2	8	
24	8:2 FTSA	8:2 Fluorotelomer sulfonic acid	39108-34-4	10	
25	10:2 FTSA	10:2 Fluorotelomer sulfonic acid	120226-60-0	12	
<b>Sulfonamides, Sulfomidoacetic acids, Sulfonamidoethanols</b>					
26	FOSA	Perfluorooctane sulfonamide	754-91-6	8	PFOSA
27	NMeFOSA	N-Methyl perfluorooctane sulfonamide	31506-32-8	9	MeFOSA
28	NEtFOSA	N-Ethyl perfluorooctane sulfonamide	4151-50-2	10	EtFOSA
29	NMeFOSAA	N-Methyl perfluorooctane sulfonamidoacetic acid	2355-31-9	11	MeFOSAA
30	NEtFOSAA	N-Ethyl perfluorooctane sulfonamidoacetic acid	2991-50-6	12	EtFOSAA





## Wisconsin PFAS Aqueous (Non-Potable Water) and Non-Aqueous Matrices Method Expectations

31	NMeFOSE	N-Methyl perfluorooctane sulfonamidoethanol	24448-09-7	11	MeFOSE
32	NEtFOSE	N-Ethyl perfluorooctane sulfonamidoethanol	1691-99-2	12	EtFOSE
<b>Replacement Chemicals</b>					
33	HFPO-DA	Hexafluoropropylene oxide dimer acid <sup>1</sup>	13252-13-6	6	PFPPrA
34	DONA	4,8-Dioxa-3H-perfluorononanoic acid <sup>2</sup>	919005-14-4	7	
35	9CI-PF3ONS	9-chlorohexadecafluoro-3-oxanonane-1-sulfonic acid <sup>3</sup>	756426-58-1	8	F-53B Major
36	11CI-PF3OUdS	11-chloroeicosafluoro-3-oxaundecane-1-sulfonic acid <sup>4</sup>	763051-92-9	10	F-53B Minor
	1 - Also referred to as "GenX"				
	2 - Also available as the ammonium salt = ADONA (Ammonium 4,8-dioxa-3H-perfluorononanoate) # 958445-44-8				
	3 - Also available as the potassium salt = Potassium, 9-chlorohexadecafluoro-3-oxanone-1-sulfonate # 73606-19-6				
	4 - Also available as the potassium salt = Potassium, 11-chloroeicosafluoro-3-oxaundecane-1-sulfonate # 83329-89-9				



## XII. Wisconsin Laboratory Accreditation Program PFAS Certification Offerings with Ions – 10.27.19

*The masses presented are expected to be used, although if other masses are used for the precursor or product ions, the reason is expected to be documented (such as interferences). If the confirmation ion is weak ( $S/N < 3$ ), it does not have to be used but instrument optimization can increase the  $S/N$ .*

#	Acronym	Name	CAS #	Precursor Ion Mass	Primary Product Ion Mass	Suggested Confirmation Product Ion Mass
<b>Carboxylic Acids</b>						
1	PFBA	Perfluorobutanoic acid	375-22-4	213	169	None
2	PFPeA	Perfluoropentanoic acid	2706-90-3	263	219	69, None
3	PFHxA	Perfluorohexanoic acid	307-24-4	313	269	119
4	PFHpA	Perfluoroheptanoic acid	375-85-9	363	319	169
5	PFOA	Perfluorooctanoic acid	335-67-1	413	369	169
6	PFNA	Perfluorononanoic acid	375-95-1	463	419	219
7	PFDA	Perfluorodecanoic acid	335-76-2	513	469	219
8	PFUnA	Perfluoroundecanoic acid	2058-94-8	563	519	269
9	PFDoA	Perfluorododecanoic acid	307-55-1	613	569, 319	569, 369, 319, 269, 169
10	PFTriA	Perfluorotridecanoic acid	72629-94-8	663	619	369, 319, 269, 169
11	PFTeA	Perfluorotetradecanoic acid	376-06-7	713	669	369, 319, 269, 169
12	PFHxDA	Perfluorohexadecanoic acid	67905-19-5	813	769	369, 319, 269, 219, 169
13	PFODA	Perfluorooctadecanoic acid	16517-11-6	913	869	369, 319, 269, 219, 169
<b>Sulfonic Acids</b>						
14	PFBS	Perfluorobutanesulfonic acid	375-73-5	299	80	99
15	PFPeS	Perfluoropentanesulfonic acid	2706-91-4	349	80	99
16	PFHxS	Perfluorohexanesulfonic acid	355-46-4	399	80	99
17	PFHpS	Perfluoroheptanesulfonic acid	375-92-8	449	99, 80	99, 80
18	PFOS	Perfluorooctanesulfonic acid	1763-23-1	499	80	99
19	PFNS	Perfluorononanesulfonic acid	68259-12-1	549	80	99
20	PFDS	Perfluorodecanesulfonic acid	335-77-3	599	99, 80	99, 80
21	PFDoS	Perfluorododecanesulfonic acid	79780-39-5	699	80	99, 62
22	4:2 FTSA	4:2 Fluorotelomer sulfonic acid	757124-72-4	327	307	81, 80
23	6:2 FTSA	6:2 Fluorotelomer sulfonic acid	27619-97-2	427	407	81, 80
24	8:2 FTSA	8:2 Fluorotelomer sulfonic acid	39108-34-4	527	507	81, 80
25	10:2 FTSA	10:2 Fluorotelomer sulfonic acid	120226-60-0	627	607	587, 81, 80





Sulfonamides, Sulfomidoacetic acids, Sulfonamidoethanols						
26	FOSA	Perfluorooctane sulfonamide	754-91-6	498	78	478, 169, None
27	NMeFOSA	N-Methyl perfluorooctane sulfonamide	31506-32-8	512	169	219
28	NEtFOSA	N-Ethyl perfluorooctane sulfonamide	4151-50-2	526	169	219
29	NMeFOSAA	N-Methyl perfluorooctane sulfonamidoacetic acid	2355-31-9	570	419	512, 483
30	NEtFOSAA	N-Ethyl perfluorooctane sulfonamidoacetic acid	2991-50-6	584	419	526, 483
31	NMeFOSE	N-Methyl perfluorooctane sulfonamidoethanol	24448-09-7	616	59	122, None
32	NEtFOSE	N-Ethyl perfluorooctane sulfonamidoethanol	1691-99-2	630	59	136, None
Replacement Chemicals						
33	HFPO-DA	Hexafluoropropylene oxide dimer acid	13252-13-6	329	285, 169	285, 169, None
34	DONA	4,8-Dioxa-3H-perfluorononanoic acid	919005-14-4	377	251	85, None
35	9Cl-PF3ONS	9-chlorohexadecafluoro-3-oxanonane-1-sulfonic acid	756426-58-1	531	351	83, None
36	11Cl-PF3OUdS	11-chloroeicosafluoro-3-oxaundecane-1-sulfonic acid	763051-92-9	631	451	99, None
NOTE: ISO 21675, SW 8327, and Wellington Laboratories provide precursor, product and confirmation ions for many of the extracted internal standards						

Mass Source
EPA 537.1
DoD QSM 5.3
Janice Willey
EPA-821-R-11-007, PFAS in Sludge/Biosolids
ISO 21675
SW 8327
Wellington Laboratories
Confirmation mass have multiple sources